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(54) **BLOOD BIOMARKERS FOR SUICIDALITY**

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Related U.S. Application Data

(57) **ABSTRACT**

(63) Continuation of application No. 14/194,024, filed on Feb. 28, 2014.

(60) Provisional application No. 61/770,696, filed on Feb. 28, 2013.

Biomarkers and methods for screening expression levels of the biomarkers for predicting and tracking suicidality, as well as for monitoring response to a treatment for suicidal risk and for determining suicidal risk as a side-effect of an antidepressant are disclosed.

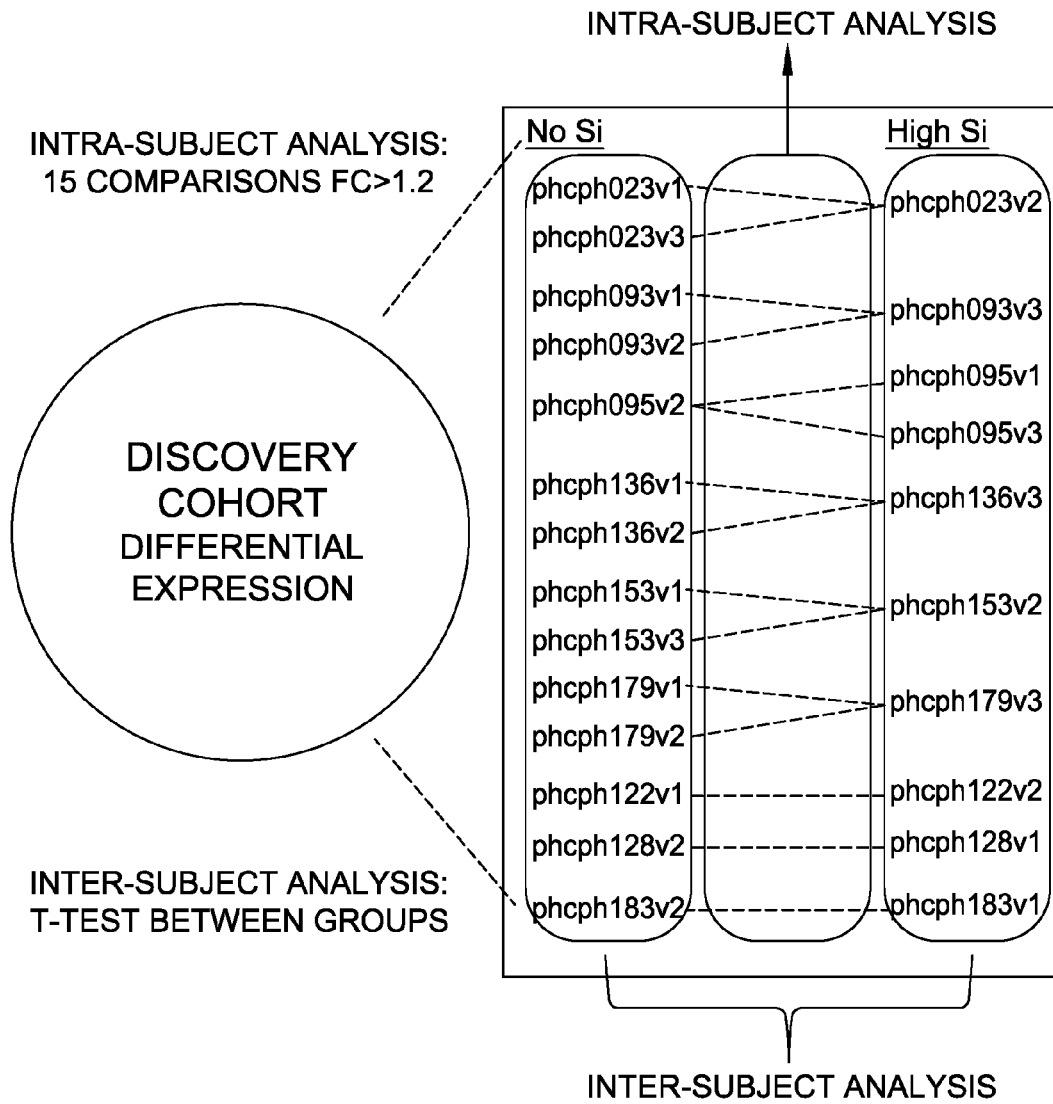


FIG. 1A

SUICIDAL IDEATION (SI)
FROM HAMILTON RATING SCALE FOR DEPRESSION (HAMD17)
No SI - SCORE OF 0; High SI - SCORE OF 2 OR ABOVE

SUICIDE

0 = ABSENT

1 = FEELS LIFE IS NOT WORTH LIVING

2 = WISHES HE WERE DEAD OR ANY THOUGHTS OF POSSIBLE
DEATH TO SELF

3 = SUICIDAL IDEAS OR GESTURE

4 = ATTEMPTS AT SUICIDE (ANY SERIOUS ATTEMPT RATES 4)

FIG. 1B

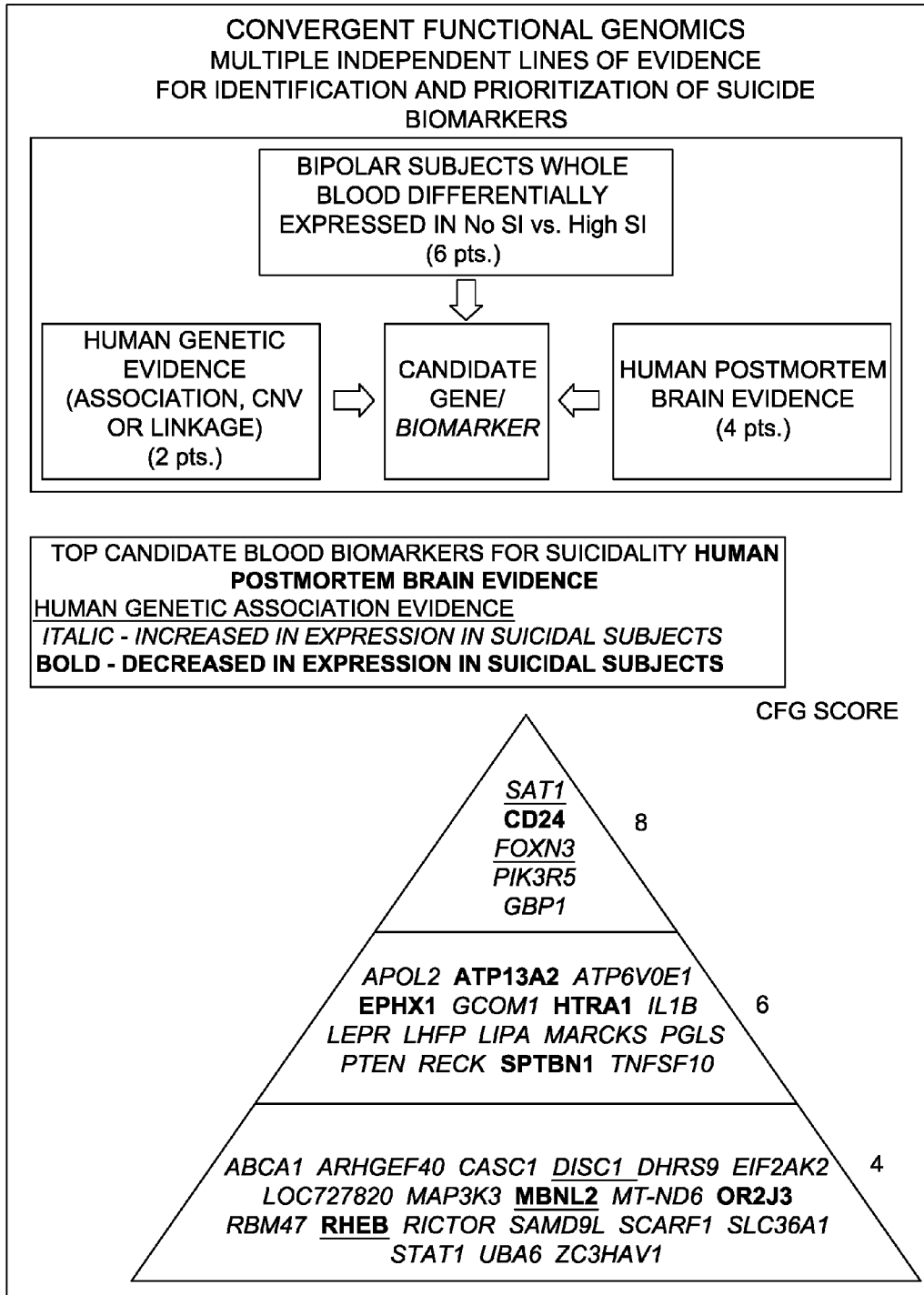


FIG. 2

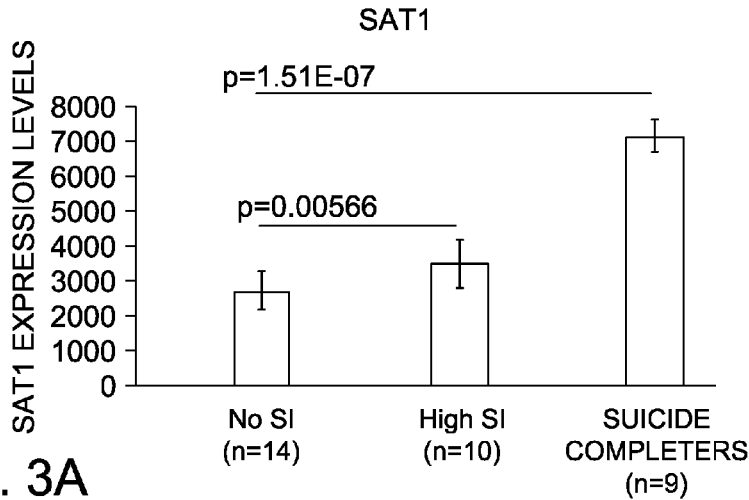


FIG. 3A

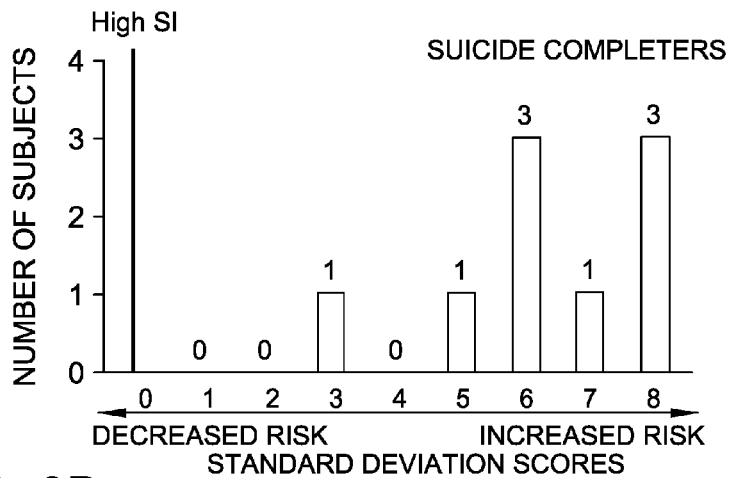


FIG. 3B

DISCOVERY COHORT	SAT1 STANDARD DEVIATION SCORES								
	0	1	2	3	4	5	6	7	8
DISCOVERY COHORT VALUES	2642.97-	3413.37-	4084.34-	4755.31-	5426.28-	6768.22-	7439.19-	8110.16-	8781.12-
DETERMINE RISK CUTOFFS	3413.37	4084.34	4755.31	5426.28	6097.25	7439.19	8110.16	8781.12	9452.1
High SI mean = 3413.37				INBR009		INBR011	INBR013 INBR017 INBR018	INBR016	INBR012 INBR014 INBR015
High SI Std. D = 670.97									

FIG. 3C

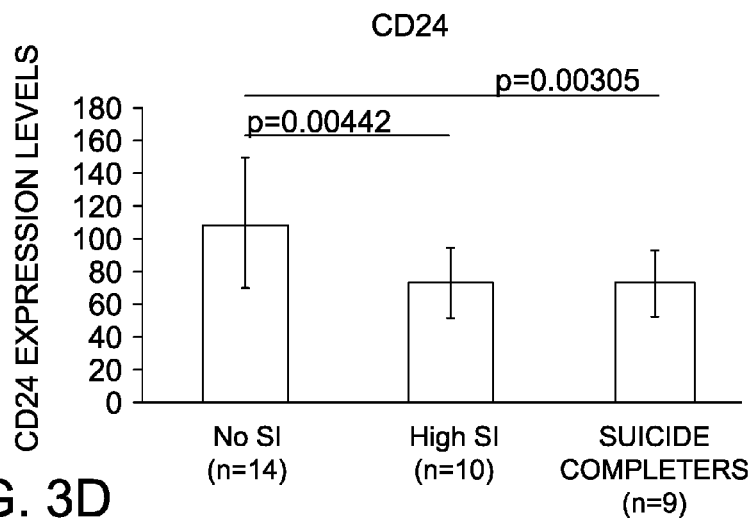


FIG. 3D

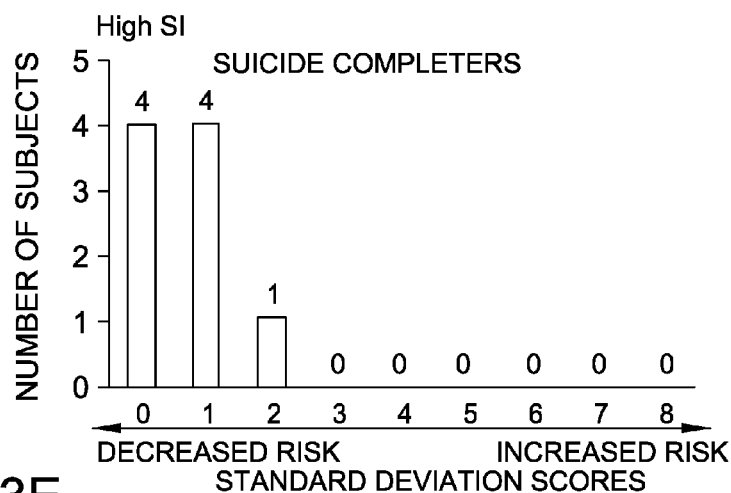


FIG. 3E

DISCOVERY COHORT	CD24 STANDARD DEVIATION SCORES									
	0	1	2	3	4	5	6	7	8	
DISCOVERY COHORT VALUES DETERMINE RISK CUTOFFS	108.63-73.01	73.01-51.53	51.53-30.01	30.01-8.58	8.58-0	0	0	0	0	
High SI mean = 73.01	INBR011	INBR009	INBR013							
High SI Std. D = 21.48	INBR014	INBR012								
	INBR016	INBR017								
	INBR018	INBR015								

FIG. 3F

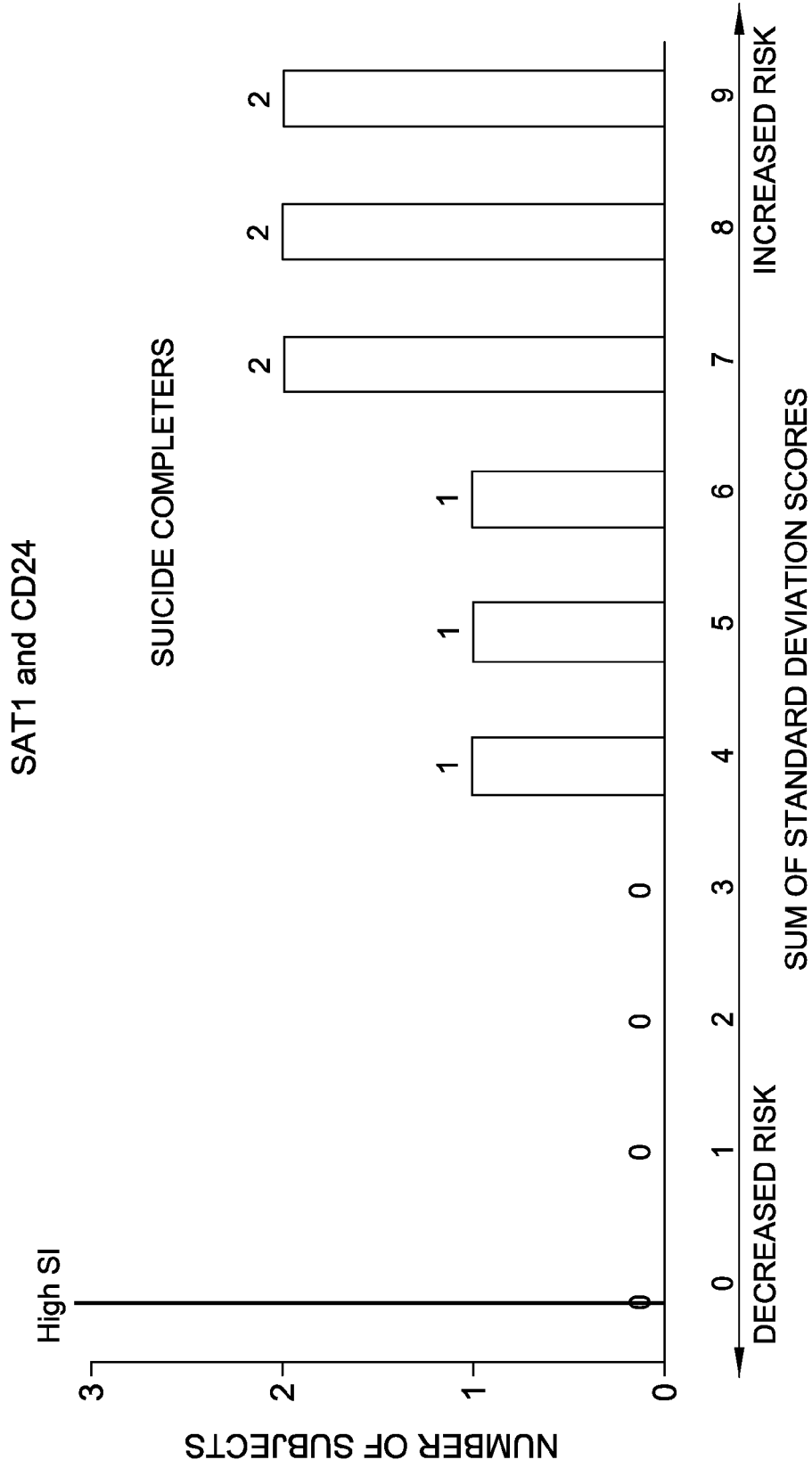


FIG. 3G

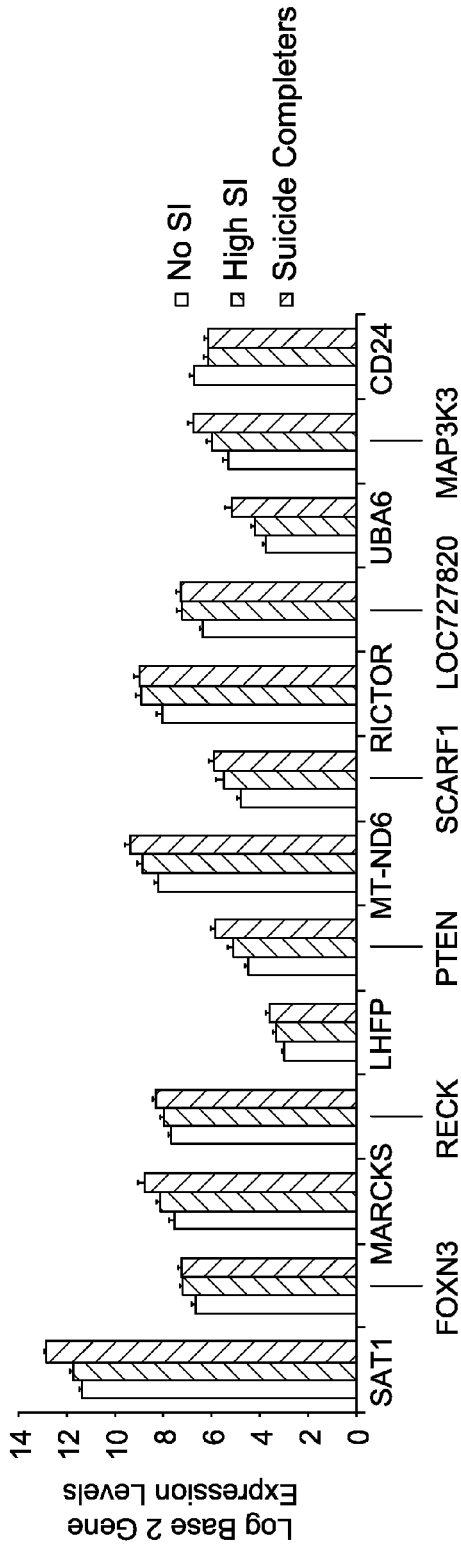


FIG. 3H

CFG Score	Gene	Direction of Change	P-Value (One-Way ANOVA)
8	SAT1		2.91E-13
4	UBA6		8.94E-05
6	MARCKS		0.000187221
6	PTEN		0.000298958
4	MT-ND6		0.000391061
4	MAP3K3		0.000777774
6	LHFPL1		0.001535921
4	LOC727820		0.003706529
8	CD24	D	0.006082658
6	RECK		0.009035235
8	FOXN3		0.010040264
4	SCARF1		0.014880001
4	RICTOR		0.040726456

FIG. 3I

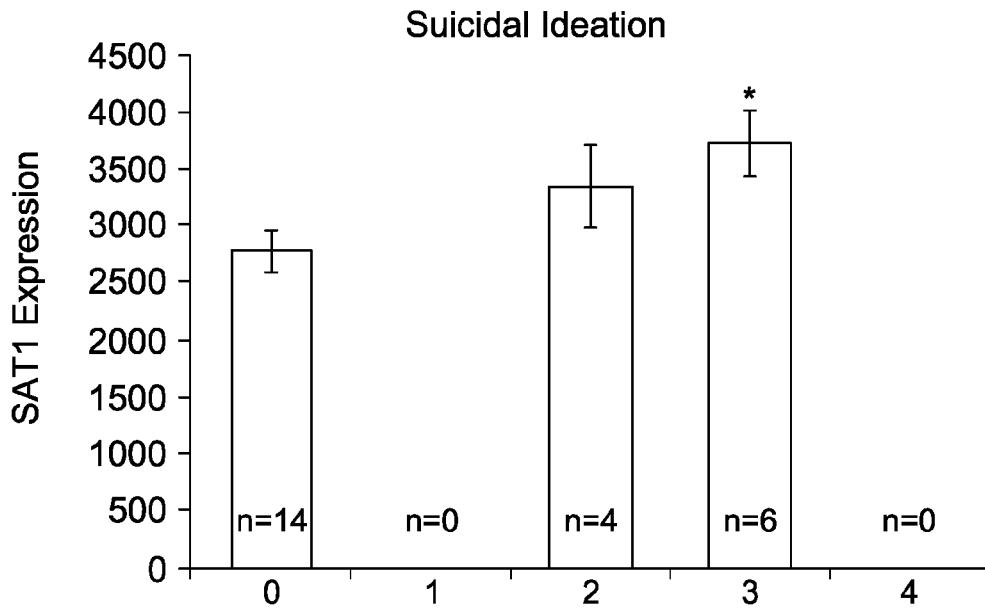


FIG. 4A

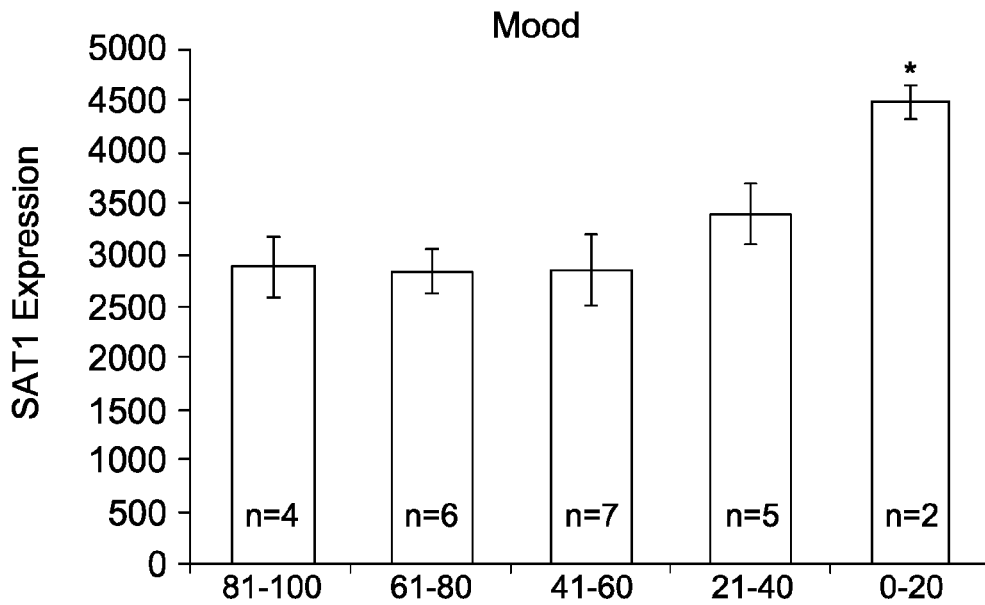


FIG. 4B

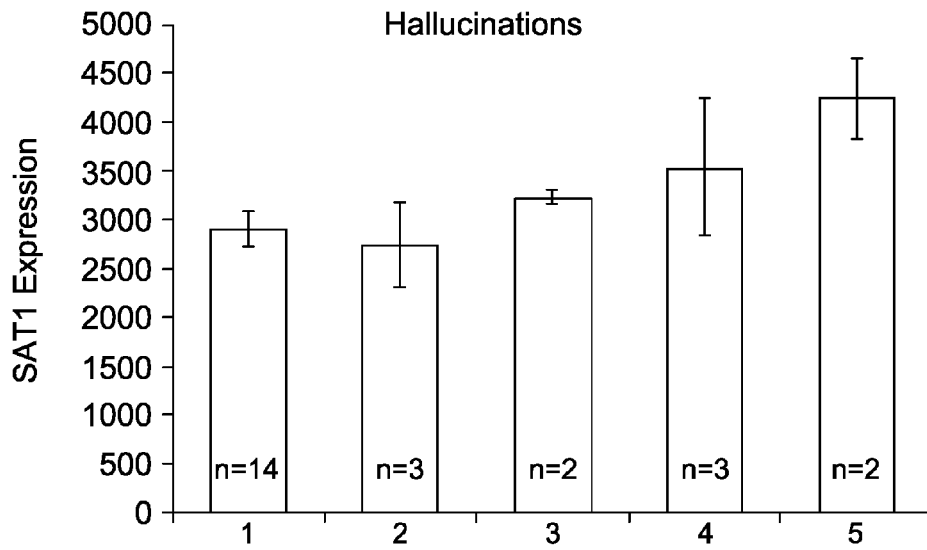


FIG. 4C

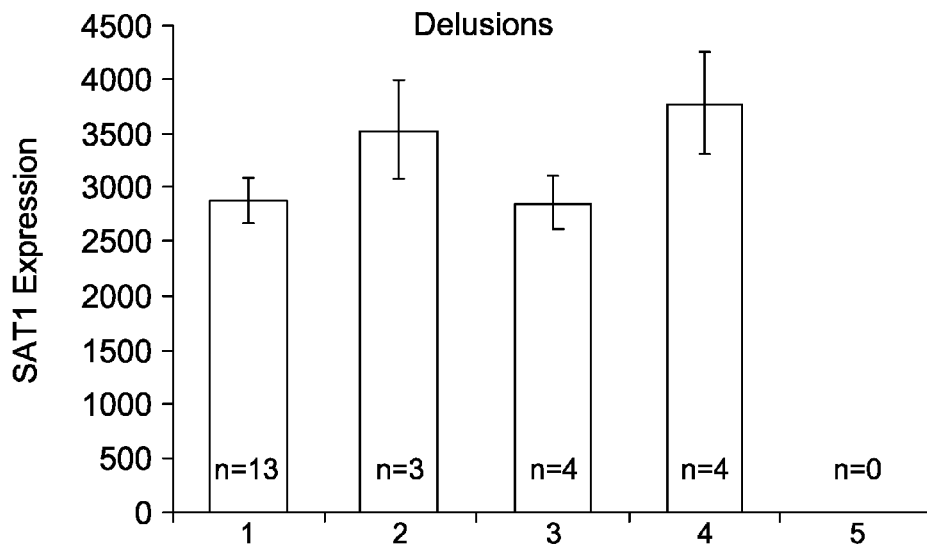


FIG. 4D

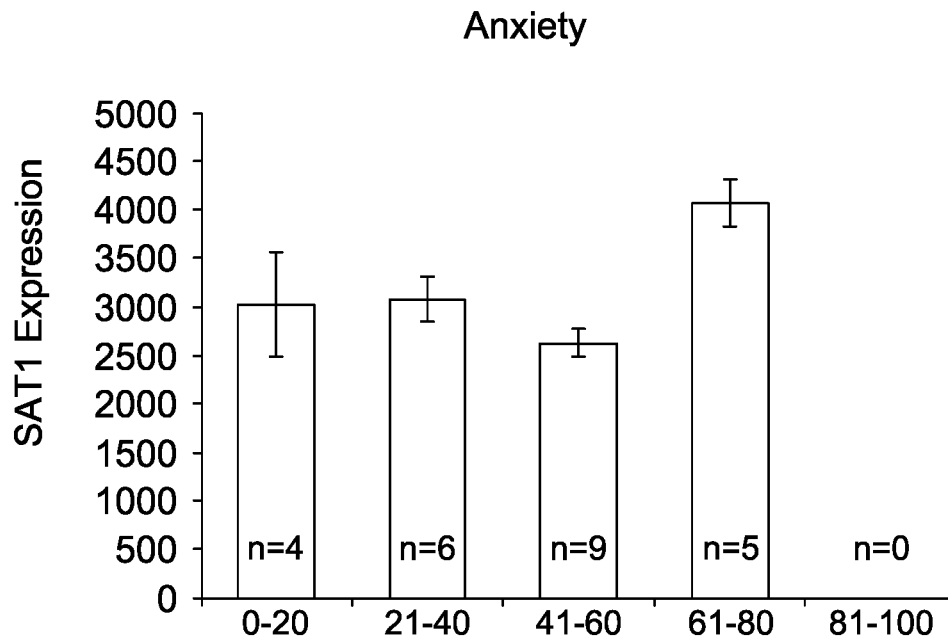


FIG. 4E

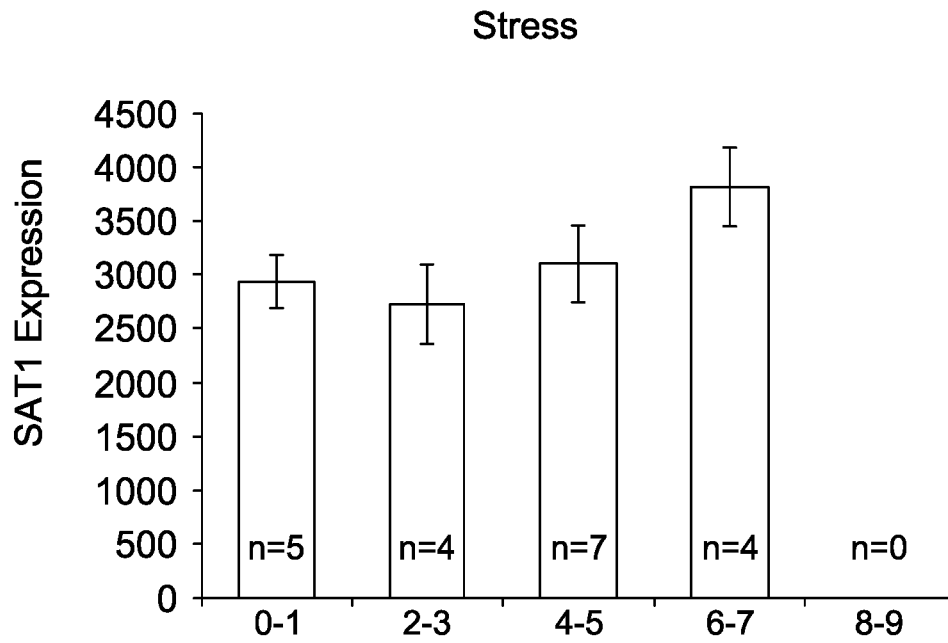


FIG. 4F

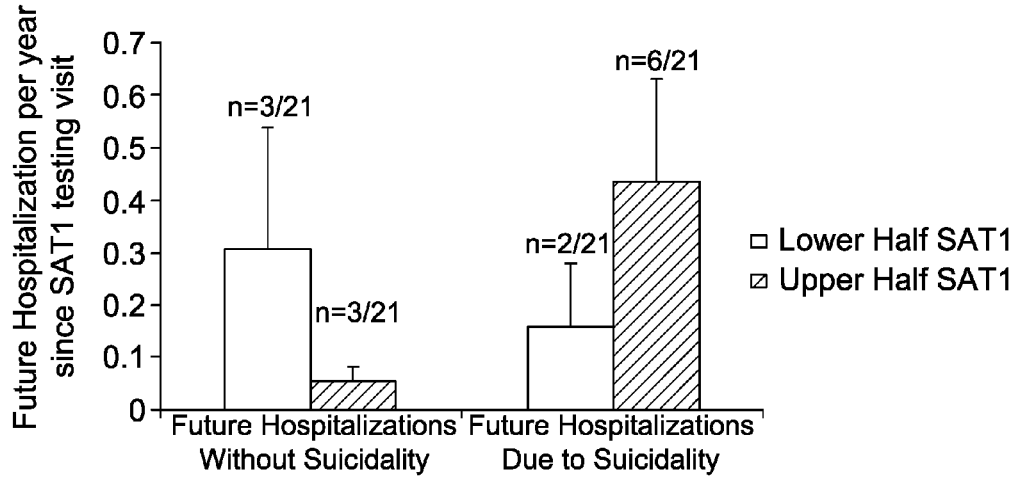


FIG. 5A

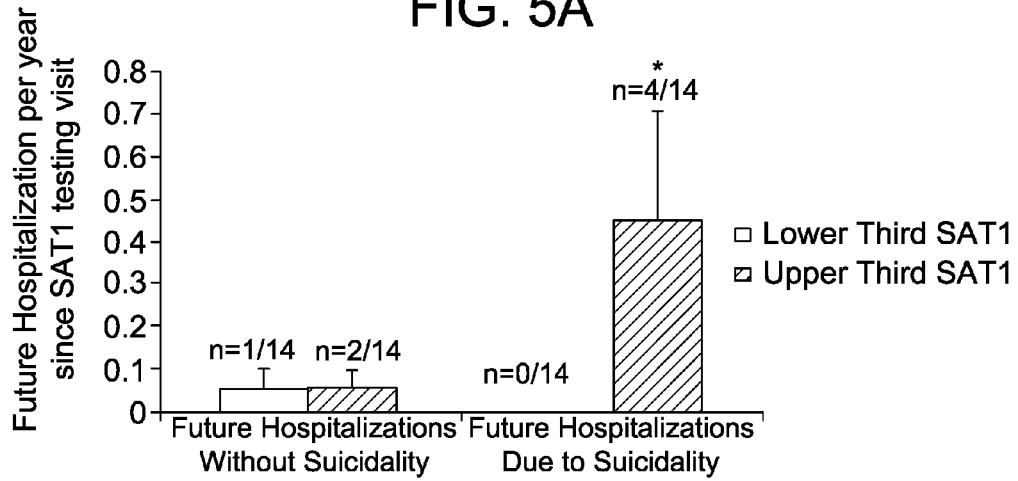


FIG. 5B

	Hospitalizations Without Suicidality t-test	Hospitalizations Due to Suicidality t-test
Upper Half SAT1 vs. Lower Half SAT1	0.14028	0.1195
Upper Third SAT1 vs. Lower Third SAT1	0.4827	0.0484

FIG. 5C

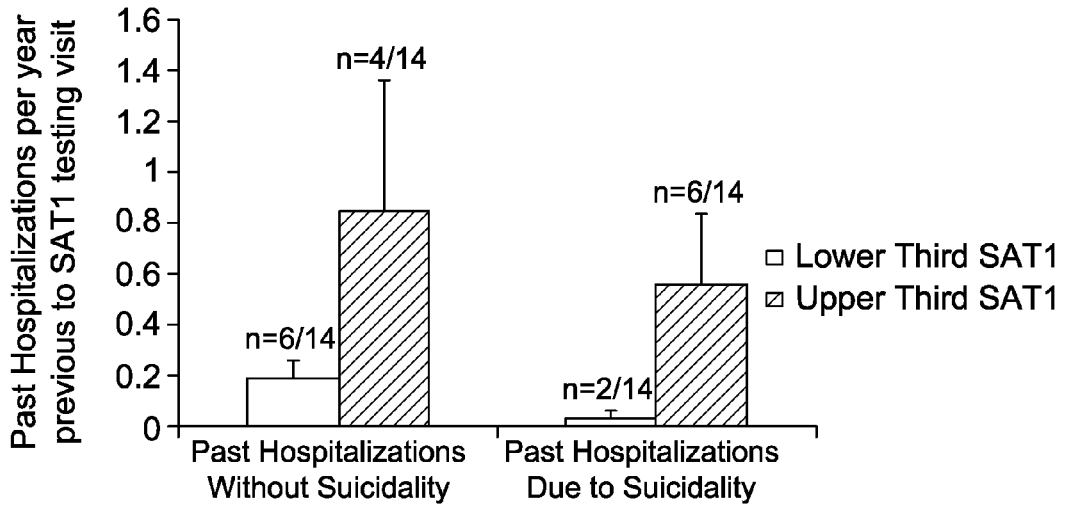


FIG. 5D

	Hospitalizations Without Suicidality t-test	Hospitalizations Due to Suicidality t-test
Bipolar Upper Third SAT1 vs. Lower Third SAT1	0.1108	0.03626

FIG. 5E

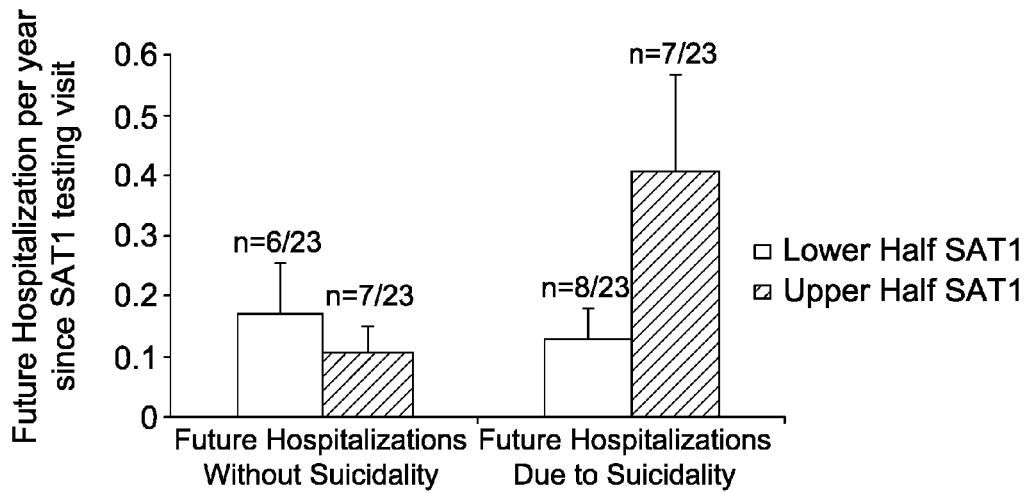


FIG. 6A

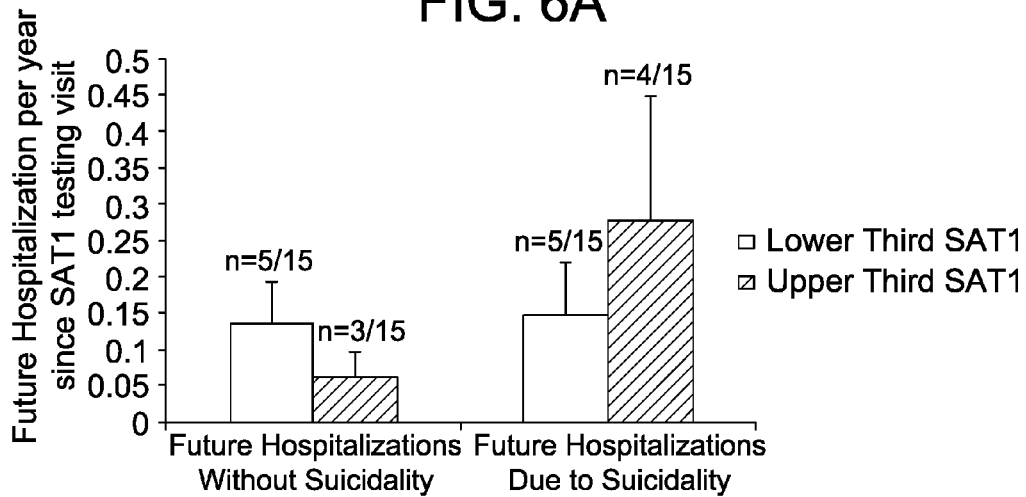


FIG. 6B

	Hospitalizations Without Suicidality t-test	Hospitalizations Due to Suicidality t-test
Upper Half SAT1 vs. Lower Half SAT1	0.2346	0.0519
Upper Third SAT1 vs. Lower Third SAT1	0.1251	0.2461

FIG. 6C

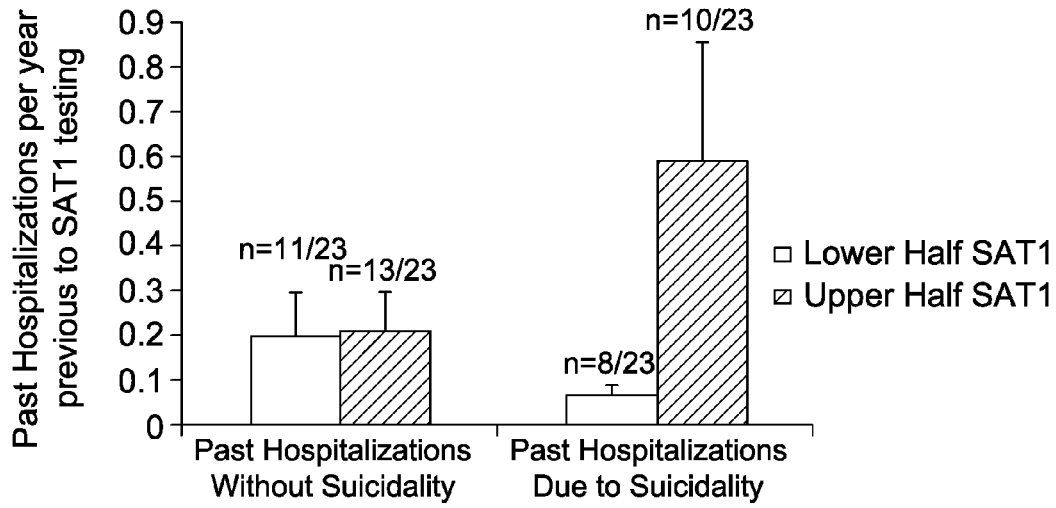


FIG. 6D

	Hospitalizations Without Suicidality t-test	Hospitalizations Due to Suicidality t-test
Psychosis Upper Half SAT1 vs. Lower Half SAT1	0.4564	0.0274

FIG. 6E

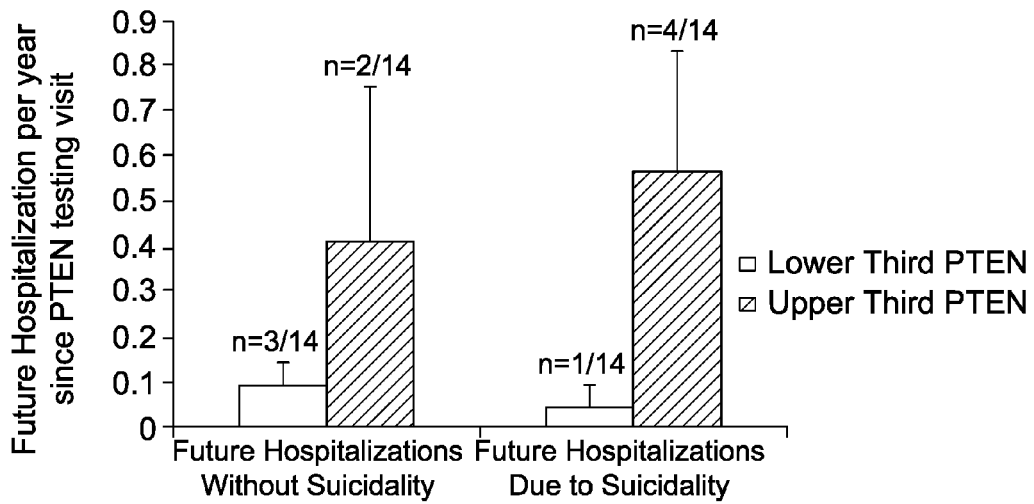


FIG. 7A

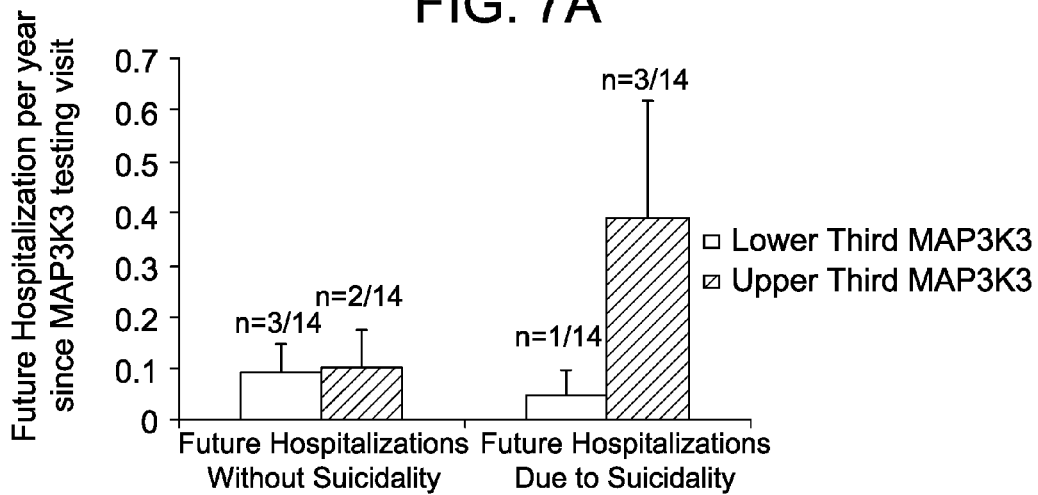


FIG. 7B

	Hospitalizations Without Suicidality t-test	Hospitalizations Due to Suicidality t-test
Upper Third PTEN vs. Lower Third PTEN	0.1856	0.0324
Upper Third MAP3K3 vs. Lower Third MAP3K3	0.4570	0.0724

FIG. 7C

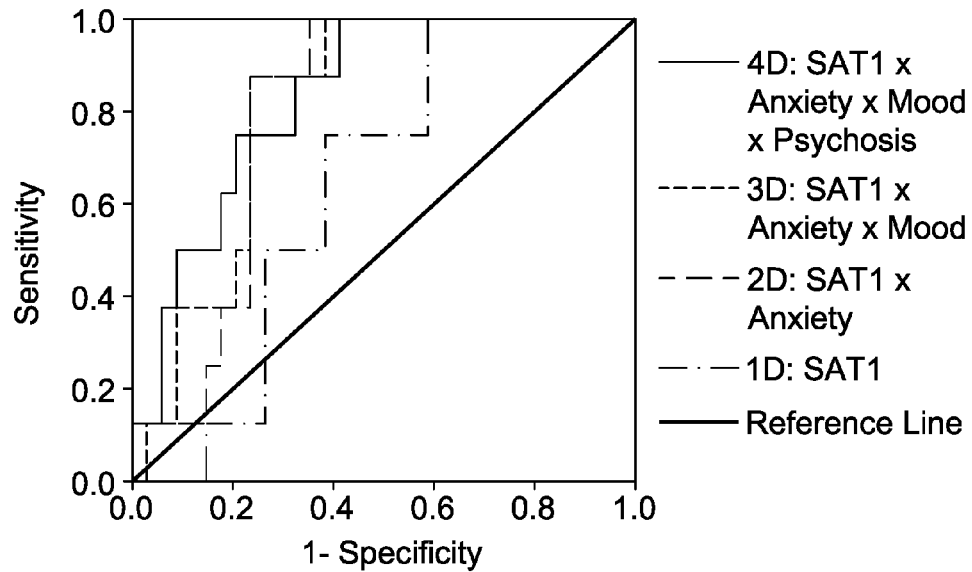


FIG. 8A

D	Test Result Variable(s)	Area Under the Curve	Std. Error	Significance	95% Confidence Interval	
					Lower Bound	Upper Bound
1D	SAT1	.640	.086	.224	.471	.808
2D	SAT1 x Anxiety	.798	.068	.009	.665	.931
3D	SAT1 x Anxiety x Mood	.813	.066	.006	.683	.942
4D	SAT1 x Anxiety x Mood x Psychosis	.835	.066	.004	.706	.964

FIG. 8B

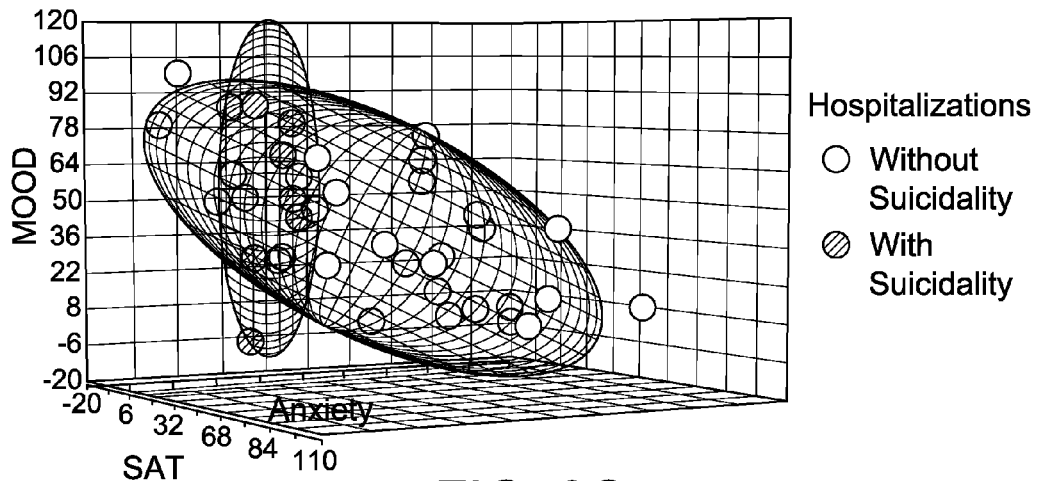


FIG. 8C

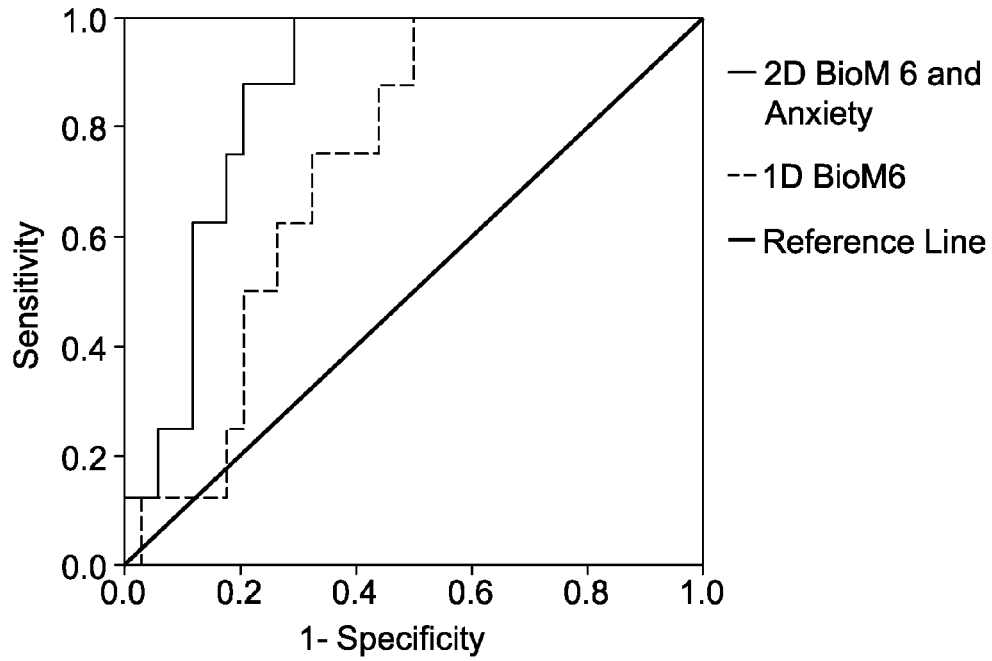


FIG. 9A

D	Test Result Variable(s)	Area Under the Curve	Std. Error	Significance	95% Confidence Interval	
					Lower Bound	Upper Bound
1D	BioM6	.732	.079	.044	.578	.886
2D	BioM6 x Anxiety	.864	.056	.002	.754	.974

FIG. 9B

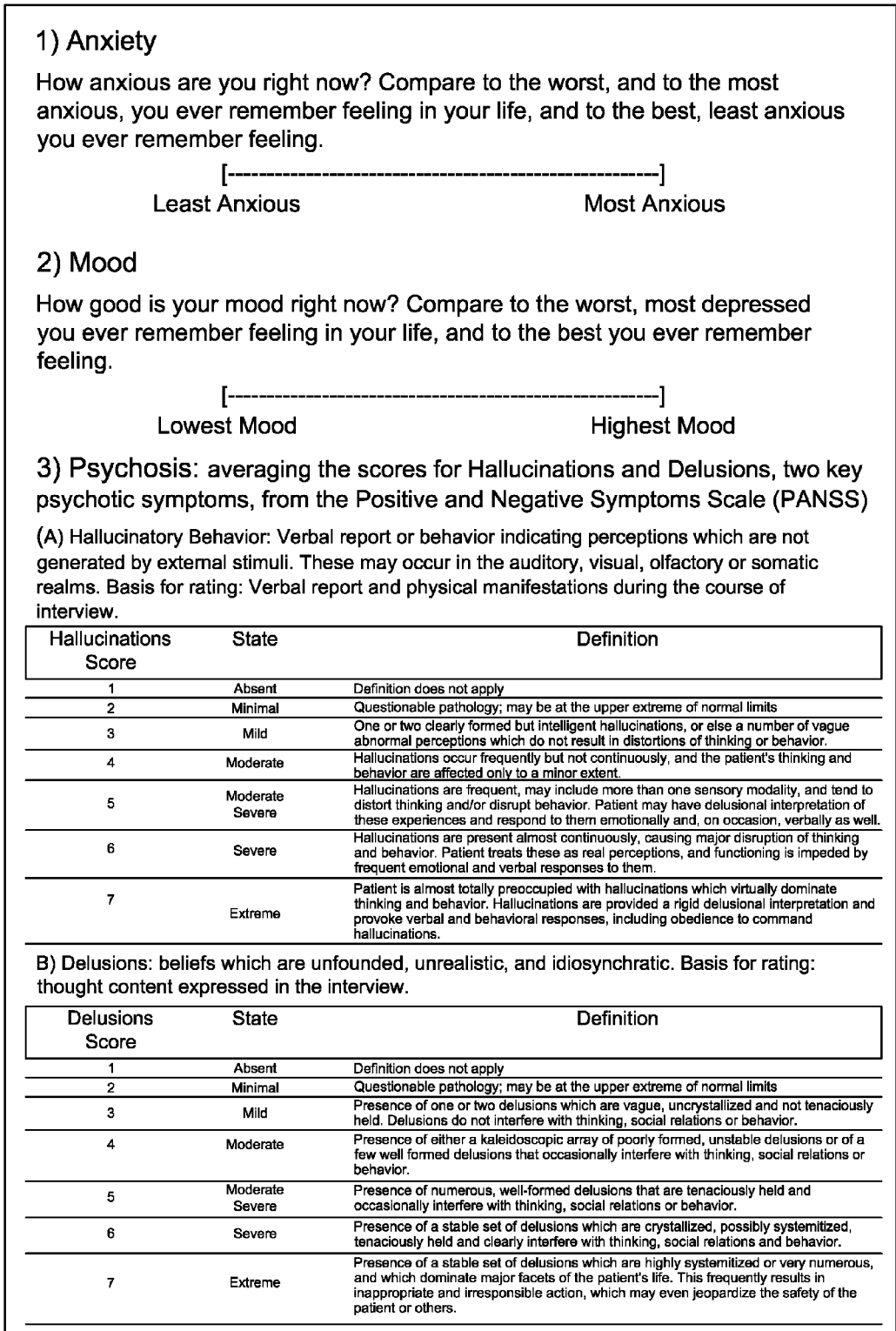


FIG. 10

BLOOD BIOMARKERS FOR SUICIDALITY

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a continuation application of U.S. application Ser. No. 14/194,024 filed on Feb. 28, 2014, which claims priority to U.S. Provisional Patent Application No. 61/770,696 filed on Feb. 28, 2013, both of which are hereby incorporated by reference in their entireties.

STATEMENT OF GOVERNMENT SUPPORT

[0002] This invention was made with government support under OD007363 awarded by the National Institutes of Health. The Government has certain rights in the invention.

BACKGROUND OF THE DISCLOSURE

[0003] The present disclosure relates generally to blood biomarkers and their use for predicting mental state, and in particular, for predicting a subject's risk of suicide (also referred to herein as "suicidality"). More particularly, the present disclosure relates to gene expression biomarkers, and to methods of screening for biomarkers, for identifying subjects who are at risk of committing suicide and methods for monitoring response to potential treatments by analyzing biomarkers.

[0004] Suicides are a leading cause of death in psychiatric patients, and in society at large. Particularly, suicide accounts for one million deaths worldwide each year. There are currently no objective tools to assess and track changes in suicidal risk without asking the subjects directly. Such tools, however, could prove substantially advantageous as the subjects at risk often choose not to share their suicidal ideation or intent with others, for fear of stigma, hospitalization, or that, in fact, their plans will be thwarted.

[0005] Conventionally, a convergence of methods assessing the subject's internal subjective feelings and thoughts, along with external, more objective, ratings of actions and behaviors, are used de facto in clinical practice, albeit not in a formalized and systematic way. Accordingly, there exists a need to develop more quantitative and objective ways for predicting and tracking suicidal states. More particularly, it would be advantageous if objective screening methods could be developed for determining expression levels of biomarkers to allow for determining suicidal risk and other psychotic depressed mood states, as well as monitoring a subject's response to treatments for lessening suicidal risk.

SUMMARY OF THE DISCLOSURE

[0006] The present disclosure relates generally to predicting and tracking suicidality. Particularly, the present disclosure is directed to screening expression levels of biomarkers for predicting and tracking suicidality, and other psychotic depressed mood states, as well as for monitoring response to a treatment for suicidal risk. In one embodiment, the screening methods are useful in determining the suicidal risk of antidepressant treatment in a subject, which has been shown to be rare, but very serious in certain situations.

[0007] Biomarkers useful for identifying subjects at risk for suicide, as well as useful for monitoring the risk of suicide following treatment have been discovered. Accordingly, the present disclosure is directed to methods of identifying a subject at risk for suicide. The present disclosure is further

directed to methods for monitoring response of a subject at risk for suicide to a treatment for suicide risk.

[0008] By monitoring and tracking changes in suicide state, the present disclosure allows for detection of an increased suicide risk prior to any suicide attempt by a subject, and further allows subjects at risk of suicide and other psychotic depressed mood states to be monitored and treated effectively. Accordingly, in another embodiment, the present disclosure relates to predicting future hospitalization for subjects being at risk for suicide and other psychotic depressed mood states such to provide sufficient monitoring and treatment to the subjects.

[0009] In one aspect, the present disclosure is directed to a method for identifying a subject at risk for suicide. The method includes obtaining a reference expression level of a blood biomarker; and determining an expression level of the blood biomarker in a sample obtained from the subject, wherein a change in the expression level of the blood biomarker in the sample obtained from the subject as compared to the reference expression level indicates a risk for suicide.

[0010] In another aspect, the present disclosure is directed to a method for monitoring response of a subject to a treatment for suicidal risk. The method includes obtaining an expression level of a biomarker from the subject; administering a treatment for suicidal risk to the subject; and determining an expression level of the biomarker in a sample obtained from the subject after the treatment is administered, wherein a change in the expression level of the biomarker in the sample obtained from the subject after the treatment is administered as compared to the expression level before administration indicates a response to the treatment.

[0011] In another aspect, the present disclosure is directed to a method for determining suicidal risk of an antidepressant, the method comprising: obtaining an expression level of a biomarker from a subject; administering an antidepressant to the subject; and determining an expression level of the biomarker in a sample obtained from the subject after the antidepressant is administered, wherein a change in the expression level of the biomarker in the sample obtained from the subject after the antidepressant is administered as compared to the expression level of the biomarker before the antidepressant is administered indicates a suicidal risk.

BRIEF DESCRIPTION OF THE DRAWINGS

[0012] The disclosure will be better understood, and features, aspects and advantages other than those set forth above will become apparent when consideration is given to the following detailed description thereof. Such detailed description makes reference to the following drawings, wherein:

[0013] FIG. 1A depicts the Discovery Cohort intra-subject and inter-subject analyses as discussed in Example 1.

[0014] FIG. 1B depicts the HAMD17 Suicidal Ideation scores as discussed in Example 1.

[0015] FIG. 2 depicts the convergent functional genomics (CFG) approach for identification and prioritization of genomic biomarkers for suicidality as discussed in Example 1.

[0016] FIGS. 3A-I depict the validation of biomarkers in the Validation Cohort (i.e., suicide completers) as discussed in Example 1.

[0017] FIGS. 4A-F depict SAT1 expression in the bipolar discovery cohort: relationship with suicidal ideation, mood, psychosis, anxiety, and stress as discussed in Example 1.

[0018] FIGS. 5A-E depict SAT1 expression levels versus subsequent hospitalizations due to suicidality as analyzed in Example 2.

[0019] FIGS. 6A-E depict SAT1 expression levels versus prediction of future hospitalizations due to suicidality as analyzed in Example 2.

[0020] FIGS. 7A-C depict expression levels of PTEN and MAP3K3 versus prediction of future hospitalizations due to suicidality as analyzed in Example 2.

[0021] FIGS. 8A-8C depict multi-dimensional prediction of future psychiatric hospitalizations due to suicidality as analyzed in Example 2. Data in each dimension was normalized to a 0-100 scale (with the mood VAS scale inverted, as the assumption was made that depressed mood states would more closely correlate with suicidality). The angle between dimensions was assumed to be 90 degrees, and a simple Pythagorean distance from origin score was calculated. The distribution of this score in the test cohort was used to generate an ROC curve for hospitalizations due to suicidality. FIG. 8A). ROC curve. FIG. 8B). Detailed results. FIG. 8C). 3 D visualization.

[0022] FIGS. 9A and 9B depict multi-dimensional prediction of future psychiatric hospitalizations due to suicidality as analyzed in Example 2. Data in each dimension was normalized to a 0-100 scale (with the mood VAS scale inverted, as the assumption was made that depressed mood states would more closely correlate with suicidality). The angle between dimensions was assumed to be 90 degrees, and a simple Pythagorean distance from origin score was calculated. The distribution of this score in the test cohort was used to generate an ROC curve for hospitalizations due to suicidality. FIG. 9A). ROC curve. FIG. 9B). Detailed results.

[0023] FIG. 10 depicts the clinical measures used in the multi-modal approach in FIGS. 8A-8C.

[0024] While the disclosure is susceptible to various modifications and alternative forms, specific embodiments thereof have been shown by way of example in the drawings and are herein described below in detail. It should be understood, however, that the description of specific embodiments is not intended to limit the disclosure to cover all modifications, equivalents and alternatives falling within the spirit and scope of the disclosure as defined by the appended claims.

DETAILED DESCRIPTION

[0025] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the disclosure belongs. Although any methods and materials similar to or equivalent to those described herein may be used in the practice or testing of the present disclosure, the preferred materials and methods are described below.

[0026] In accordance with the present disclosure, biomarkers useful for objectively identifying subjects at risk for suicide, as well as for monitoring the risk of suicide following treatment and determining the risk of suicide following administration of antidepressants have been discovered. In one aspect, the present disclosure is directed to a method for identifying a subject at risk for suicide. The method includes obtaining a reference expression level of a blood biomarker; and determining an expression level of the blood biomarker in a sample obtained from the subject. A change in the expression level of the blood biomarker in the sample obtained from the subject as compared to the reference expression level indicates a risk for suicide. In some embodiments, the meth-

ods further include obtaining clinical risk factor information and clinical scale data such as for anxiety, mood and/or psychosis from the subject in addition to obtaining blood biomarker expression level in a sample obtained from the subject. This combined clinical data and blood biomarker expression level can further improve predictability of the risk of suicide as shown in FIGS. 8A-8C and 9A-9B.

[0027] As used herein, “a subject at risk for suicide” refers to a subject diagnosed by one skilled in the art such as, for example, a clinician, using established protocols and methods for diagnosing suicidality. Such methods can include, for example, rigorous clinical interview using clinical standards for assessing and diagnosing whether a subject is at risk for suicide. Suicidality diagnosis can be established using, for example, questionnaires to identify suicidal ideation. Diagnosis can include diagnostic assessment using psychiatric rating scales including, for example, the Hamilton Rating Scale for Depression (HAMD-17), which includes a suicidal ideation rating item, Beck Scale for suicide ideation, Columbia Suicide Severity Rating Scale, The Kessler Psychological Distress Scale, and combinations thereof.

[0028] Particularly suitable subjects are humans. Suitable subjects can also be experimental animals such as, for example, monkeys and rodents, that display a behavioral phenotype associated with suicide, for example, a mood disorder or psychosis.

[0029] As used herein, “a reference expression level of a biomarker” refers to the expression level of a biomarker established for a subject with no suicidal ideation, expression level of a biomarker in a normal/healthy subject with no suicidal ideation as determined by one skilled in the art using established methods as described herein, and/or a known expression level of a biomarker obtained from literature. As known by those skilled in the art, “suicidal ideation” refers to thoughts, feelings, intent, external actions and behaviors about completing suicide. Suicidal ideation can vary from fleeting thoughts to unsuccessful attempts.

[0030] As used herein, “expression level of a biomarker” refers to the process by which a gene product is synthesized from a gene encoding the biomarker as known by those skilled in the art. The gene product can be, for example, RNA (ribonucleic acid) and protein. Expression level can be quantitatively measured by methods known by those skilled in the art such as, for example, northern blotting, amplification, polymerase chain reaction, microarray analysis, tag-based technologies (e.g., serial analysis of gene expression and next generation sequencing such as whole transcriptome shotgun sequencing or RNA-Seq), Western blotting, and combinations thereof.

[0031] Suitable biomarkers found to have a change in expression level include, for example, spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H+ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversal-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange fac-

tor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAMD9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1), CD24, ATP13A2, EPHX1, HTRA1, SPTBN1, MBNL2, OR2J3, RHEB, DBP, and combination thereof. Particularly suitable biomarkers include SAT1, MARCKS, PTEN, MAP3K3, and combinations thereof.

[0032] As used herein, a “change” in the expression level of the biomarker refers to an increase or a decrease of by about 1.2-fold or greater in the expression level of the biomarker as determined in a sample obtained from the subject as compared to the reference expression level of the biomarker. In one embodiment, the change in expression level is an increase or decrease by about 1.2 fold.

[0033] In one embodiment, the expression level of the blood biomarker in the sample obtained from the subject is increased as compared to the reference expression level of the biomarker. It has been found that an increase in the expression level of particular blood biomarkers in the sample obtained from the subject as compared to the reference expression level of the biomarker indicates a risk for suicide. Suitable biomarkers that indicate a risk for suicide when the expression level increases can be, for example, spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H+ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAMD9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator

of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2), and combinations thereof. See, Table 5 for a list of biomarkers identified as showing an increase in expression level.

[0034] In another embodiment, the expression level of the blood biomarker in the sample obtained from the subject is decreased as compared to the reference expression level of the biomarker. Suitable biomarkers that indicate a risk for suicide when the expression level decreases as compared to the reference expression level have been found to include, for example, cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof. See, Table 5 for a list of biomarkers identified as showing a decrease in expression level.

[0035] In another embodiment, the method includes determining the expression level of a blood biomarker in the sample obtained from the subject that is increased as compared to the reference expression level of the biomarker and determining the expression level of the blood biomarker in the sample obtained from the subject that is decreased as compared to the reference expression level of the biomarker. For example, spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H+ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAMD9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-

like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2), and combinations thereof in the blood sample of the subject can be increased as compared to the reference expression level, and cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof in the blood sample of the subject can be decreased as compared to the reference expression level to indicate an increase in the risk of suicide in a subject.

[0036] A particularly suitable sample for which the expression level of a biomarker is determined can be, for example, blood, including whole blood, leukocytes, and megakaryocytes. Other suitable samples for which the expression level of a biomarker is determined can be, for example, brain, cerebrospinal fluid, olfactory epithelium cells, fibroblasts from skin biopsies, induced pluripotent stem cells, and neuronal-like cells derived therefrom.

[0037] While described herein as a change in expression level, in some embodiments, particular levels of one or more of the above-described biomarkers can be useful for objectively identifying subjects at risk for future suicide. For example, it has been found that levels of SAT1 at 2500 Affymetrix microarray fluorescence intensity units (AU) or greater, including 2600 AU or greater, including 2700 AU or greater, including 2800 AU or greater, including 2900 AU or greater, and including 3000 AU or greater, have been found to be at increased risk for future suicide.

[0038] In another aspect, the present disclosure is directed to a method for monitoring response of a subject to a treatment for suicidal risk. As used herein, "treatment for suicidal risk" refers to a drug, nutritional, pharmaceutical, or the like, and combinations thereof that can modify the likelihood of a subject attempting and/or completing suicide. The method includes obtaining an expression level of a biomarker; administering a treatment for suicidal risk to the subject; and determining the expression level of the biomarker in a sample obtained from the subject after the treatment is administered, wherein a change in the expression level of the biomarker in the sample obtained from the subject after the treatment is administered as compared to the expression level of the biomarker before the treatment is administered indicates a response to the treatment.

[0039] Administration of the treatment can be by any suitable method known by those skilled in the art such as, for example, topical administration, enteral administration and parenteral administration. Suitable methods of administration can be, for example, transdermal administration, oral administration, and injection.

[0040] Suitable treatments for suicidal risk can be, for example, clozapine, omega-3 fatty acids (e.g., docosahexaenoic acid (DHA)), lithium, IL-1 trap, canakinumab, nicorandil, amiodarone, arsenic trioxide, vemurafenib, elsamitrucin, T 0128, CT-2106, BN80927, tafluposide, TAS-

103, beta-lapachone, irinotecan, topo tecan, 9-amino-20-camptothecin, rubitecan, gimimatecan, karenitecin, and combinations thereof.

[0041] Response to the treatment can be a decrease in the expression level of a biomarker after treatment. Biomarkers for which a decrease in the expression level of the biomarker indicates a response to the treatment can be, for example, spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H⁺ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAMD9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2), and combinations thereof.

[0042] Response to the treatment can alternatively be an increase in the expression level of a biomarker after treatment. Biomarkers for which an increase in the expression level of the biomarker indicates a response to the treatment can be, for example, small cell lung carcinoma cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein and combinations thereof.

[0043] Response to the treatment can be a decrease in the expression level of a first biomarker and an increase in a second biomarker. The first biomarker can be, for example, spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H⁺ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase

(PGLS); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAMD9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); and combinations thereof. The second biomarker can be, for example, cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, interleukin 1 beta (IL1B), phosphatase and tensin homolog (PTEN), promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof. See, Table 5.

[0044] In another aspect, the present disclosure is directed to a method for determining suicidal risk as a side-effect of an antidepressant. The method includes obtaining an expression level of a biomarker from the subject; administering an antidepressant to the subject; and determining an expression level of the biomarker in a sample obtained from the subject after the antidepressant is administered. A change in the expression level of the biomarker in the sample obtained from the subject after the antidepressant is administered as compared to the expression level of the biomarker before the antidepressant is administered indicates suicidal risk as a side-effect of the antidepressant.

[0045] It is known that suicide risk is a rare, but very serious side-effect of some drugs. Upon initiation of antidepressant therapy, subjects can sometimes experience a sudden onset of suicidal ideation (e.g., suicidal thoughts and behaviors) that accompanies treatment. Subjects can become suicidal in the first weeks of treatment, upon a dosage change and/or a combination thereof. This has caused the U.S. Food and Drug Administration to require manufacturers to place explicit warnings on the label of the drug stating that its use may cause a risk of suicide.

[0046] Suitable antidepressants can be, for example, bupropion, citalopram, escitalopram, fluoxetine, fluvoxamine, mirtazapine, nefazodone, paroxetine, sertraline, and venlafaxine.

[0047] Suitable biomarkers can be, for example, spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phos-

phoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H⁺ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAMD9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2); cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof. Particularly suitable biomarkers include SAT1, MARCKS, PTEN, MAP3K3, and combinations thereof.

[0048] In yet another aspect, the present disclosure is directed to a method of predicting hospitalization of a subject at risk of suicide. The method includes obtaining a first expression level of a blood biomarker in an initial sample obtained from the subject; and determining a second expression level of the blood biomarker in a subsequent sample obtained from the subject, wherein an increase in the expression level of the blood biomarker in the subsequent sample obtained from the subject as compared to the expression level of the initial sample indicates a higher risk of future hospitalizations due to suicidality.

[0049] Suitable biomarkers can be, for example, spermidine/spermine N1-acetyltransferase 1 (SAT1); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); mitogen-activated protein kinase kinase 3 (MAP3K3); and combinations thereof.

EXAMPLES

Example 1

Materials and Methods

[0050] In this Example, whole-genome gene expression profiling of blood samples was conducted to identify blood gene expression biomarkers for suicidality.

[0051] Human Subjects.

[0052] Male Caucasian subjects diagnosed with bipolar disorder (“Discovery Cohort”) were evaluated that had a diametrical change in suicidal ideation scores from no suicidal ideation to high suicidal ideation from visit to visit. The subjects were limited to minimize any potential gender-related and ethnicity-related state effects on gene expression. A demographic breakdown of the Discovery Cohort subjects is shown in Table 1A.

[0053] A “Validation Cohort”, in which the top biomarker findings from the Discovery Cohort testing were evaluated, consisted of an age-matched cohort of 9 male suicide completers obtained through the Marion County Coroner’s Office (8 Caucasians, 1 African-American) (Table 1B). The subjects in the Validation Cohort were required to have a last observed alive post-mortem interval of 24 hours or less, and had to have completed suicide by means other than overdose, which could affect gene expression.

TABLE 1

Demographics (1) Detailed. (2) Aggregate. Diagnosis established by comprehensive structured clinical interview (DIGS). NOS—not otherwise specified. Suicidal Ideation question is from the Hamilton Rating Scale for Depression obtained at the time of the blood draw for each subject.					
(1) A. Discovery Cohort					
SubjectID-Visit	Diagnosis	Age	Gender	Ethnicity	Suicidal Ideation
phchp023v1	Bipolar Disorder NOS	52	M	Caucasian	0
phchp023v2	Bipolar Disorder NOS	52	M	Caucasian	3
phchp023v3	Bipolar Disorder NOS	52	M	Caucasian	0
phchp093v1	Bipolar I Disorder	51	M	Caucasian	0
phchp093v2	Bipolar I Disorder	51	M	Caucasian	0
phchp093v3	Bipolar I Disorder	52	M	Caucasian	3
phchp095v1	Bipolar I Disorder	28	M	Caucasian	3
phchp095v2	Bipolar I Disorder	29	M	Caucasian	0
phchp095v3	Bipolar I Disorder	29	M	Caucasian	2
phchp122v1	Bipolar Disorder NOS	51	M	Caucasian	0
phchp122v2	Bipolar Disorder NOS	51	M	Caucasian	2
phchp128v1	Bipolar I Disorder	45	M	Caucasian	2
phchp128v2	Bipolar I Disorder	45	M	Caucasian	0
phchp136v1	Bipolar I Disorder	41	M	Caucasian	0
phchp136v2	Bipolar I Disorder	41	M	Caucasian	0
phchp136v3	Bipolar I Disorder	41	M	Caucasian	3
phchp153v1	Bipolar II Disorder	55	M	Caucasian	0
phchp153v2	Bipolar II Disorder	55	M	Caucasian	2
phchp153v3	Bipolar II Disorder	56	M	Caucasian	0
phchp179v1	Bipolar Disorder NOS	36	M	Caucasian	0
phchp179v2	Bipolar Disorder NOS	37	M	Caucasian	0
phchp179v3	Bipolar Disorder NOS	37	M	Caucasian	3
phchp183v1	Bipolar I Disorder	48	M	Caucasian	3
phchp183v2	Bipolar I Disorder	48	M	Caucasian	0

TABLE 1-continued

Demographics (1) Detailed. (2) Aggregate. Diagnosis established by comprehensive structured clinical interview (DIGS). NOS—not otherwise specified. Suicidal Ideation question is from the Hamilton Rating Scale for Depression obtained at the time of the blood draw for each subject.

B. Validation Cohort

SubjectID	Psychiatric Diagnosis	Age	Gender	Ethnicity	Suicide
INBR009	Bipolar/ Schizophrenia	59	M	Caucasian	Hanging
INBR011	Depression/ADHD	26	M	Caucasian	GSW to chest
INBR012	Unknown	39	M	Caucasian	GSW to head
INBR013	Depression	68	M	African-American	GSW to mouth
INBR014	None	27	M	Caucasian	Hanging
INBR015	None	40	M	Caucasian	Hanging
INBR016	Anxiety/TBI	68	M	Caucasian	GSW to head
INBR017	Depression	56	M	Caucasian	GSW to chest
INBR018	None	65	M	Caucasian	Slit wrist

(2)

Discovery Cohort	Suicidal Ideation		
	No SI (0)	High SI (2-4)	Overall
SI (score)			
Number of subjects (number of chips)	9(14)	9(10)	9(24)
Age mean years (SD)	46.1 (8.1)	43.8 (9.7)	45.1 (8.7)
range	29-56	28-55	28-56
Ethnicity # subjects (Caucasian/African-American)	(9/0)	(9/0)	(9/0)

Test Cohort	Suicide Completers
Number of subjects (number of chips)	9(9)
Age mean years (SD)	49.8 (17)
range	26-68
Ethnicity # subjects (Caucasian/African-American)	(8/1)

[0054] The Discovery Cohort subjects were on a variety of different psychiatric medications, including mood stabilizers, antidepressants, antipsychotics, benzodiazepines, and others as listed in Table 2A (Table 2B provides toxicology for subjects in the coroner’s office test cohort-suicide completers). Medications can have a strong influence on gene expression. However, this Example tested differentially expressed genes based at on intra-subject analyses, which factor out not only genetic background, effects but also medication effects. Moreover, there was no consistent pattern found in any particular type of medication, or between any change in medications and suicidal ideation in the rare instances where there were changes in medications between visits. Subjects were excluded, however, if they had significant acute medical or neurological illnesses, or had evidence of active substance abuse or dependence.

TABLE 2A

Psychiatric medications of Discovery Cohort subjects.

SubjectID-Visit	Psychiatric Medications
phchp023v1	FLEXARIL 10 MG FOR SLEEP PRN LAMOTRIGINE 200 MG ZIPRASIDONE 60 MG
v2	FLEXARIL 10 MG FOR SLEEP PRN LAMOTRIGINE 200 MG ZIPRASIDONE 60 MG
v3	FLEXARIL 10 MG FOR SLEEP PRN LAMOTRIGINE 200 MG ZIPRASIDONE 60 MG
Phchp093v1	CITALOPRAM HYDROBROMIDE 40 MG TAB TAKE ONE-HALF TABLET ORALLY EVERY DAY VALPROIC ACID 500 MG 24 HR (ER) SA TAB TAKE THREE TABLETS ORALLY AT BEDTIME

TABLE 2A-continued

Psychiatric medications of Discovery Cohort subjects.	
SubjectID-Visit	Psychiatric Medications
v2	<p>QUETIAPINE FUMARATE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>GABAPENTIN 300 MG CAP TAKE ONE CAPSULE ORALLY AT BEDTIME FOR 3 DAYS, THEN TAKE ONE CAPSULE, TWICE A DAY</p> <p>QUETIAPINE FUMARATE 25 MG TAB TAKE ONE TABLET ORALLY EVERY DAY AS NEEDED</p> <p>CITALOPRAM HYDROBROMIDE 40 MG TAB TAKE ONE-HALF TABLET ORALLY EVERY DAY</p> <p>VALPROIC ACID 500 MG 24 HR (ER) SA TAB TAKE THREE TABLETS ORALLY AT BEDTIME</p> <p>DOXEPIN HCL 10 MG CAP TAKE ONE CAPSULE ORALLY AT BEDTIME</p> <p>GABAPENTIN 300 MG CAP TAKE TWO CAPSULES ORALLY TWICE A DAY AND TAKE THREE CAPSULES AT BEDTIME</p> <p>QUETIAPINE FUMARATE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>QUETIAPINE FUMARATE 25 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p>
v3	<p>CITALOPRAM HYDROBROMIDE 40 MG TAB TAKE ONE-HALF TABLET ORALLY EVERY DAY</p> <p>VALPROIC ACID 500 MG 24 HR (ER) SA TAB TAKE THREE TABLETS ORALLY AT BEDTIME</p> <p>DOXEPIN HCL 10 MG CAP TAKE ONE CAPSULE OLLY AT BEDTIME</p> <p>GABAPENTIN 300 MG CAP TAKE TWO CAPSULES ORALLY TWICE A DAY WITH FOOD</p> <p>QUETIAPINE FUMARATE 100 MG TAB TAKE ONE TABLET ORALLY PENDING AT BEDTIME</p> <p>QUETIAPINE FUMARATE 25 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p>
Phchp095v1	<p>VALPROIC ACID 250 MG 24 HR (ER) SA TAB TAKE SEVEN TABLETS ORALLY AT BEDTIME</p> <p>RISPERIDONE 2 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p> <p>SERTRALILNE HCL 100 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p>
v2	<p>VALPROIC ACID 250 MG 24 HR (ER) SA TAB TAKE SEVEN TABLETS ORALLY AT BEDTIME</p> <p>RISPERIDONE 2 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p> <p>SERTRALINE HCL 100 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p>
v3	<p>BENZTROPINE MESYLATE ORAL 1 MG TAB TAKE ONE TABLET ORALLY TWICE A DAY</p> <p>TRAZODONE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>RISPERIDONE 4 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p> <p>LORAZEPAM INJ IM Q4H PRN 2 MG/1 ML</p> <p>LORAZEPAM TAB PO Q6H PRN 2 MG</p>
Phchp122v1	<p>AMITRIPTYLINE HCL 10 MG TAB TAKE ONE TABLET ORALLY THREE TIMES DAILY AT 10AM, 2PM AND 10PM</p> <p>LEVETIRACETAM 500 MG TAB TAKE ONE TABLET ORALLY TWICE A DAY</p> <p>LORAZEPAM 0.5 MG TAB TAKE 1 TABLET ORALLY TWICE A DAY</p> <p>LUBRICATING (PF) OPH OINT APPLY 1/2 INCH RIBBON IN BOTH EYES AT BEDTIME</p> <p>RISPERIDONE 4 MG TAB TAKE ONE-HALF TABLET ORALLY AT BEDTIME</p> <p>TOPIRAMATE 25 MG TAB TAKE ONE TABLET ORALLY TWICE A DAY; INCREASE AS DIRECTED TO TWO TABLETS TWICE A DAY</p> <p>TRAZODONE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME AS NEEDED FOR INSOMNIA</p>

TABLE 2A-continued

Psychiatric medications of Discovery Cohort subjects.	
SubjectID-Visit	Psychiatric Medications
v2	<p>VALPROIC ACID 500 MG 24 HR (ER) SA TAB TAKE TWO TABLETS ORALLY AT BEDTIME</p> <p>LORAZEPAM 1 MG TAB TAKE TWO TABLETS ORALLY AT BEDTIME AS NEEDED FOR INSOMNIA</p> <p>MIRTAZAPINE 30 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>PRazosin 2 MG CAP TAKE ONE CAPSULE ORALLY TWICE A DAY. TAKE SECOND DOSE AT BEDTIME.</p> <p>VENLAFAXINE HCL 150 MG 24 HR SA TAB TAKE ONE TABLET ORALLY TWICE A DAY (BREAKFAST AND LUNCH)</p> <p>ZIPRASIDON 80 MG CAP TAKE TWO CAPSULES ORALLY EVERY EVENING WITH DINNER</p>
phchp128v1	<p>DISULFIRAM 250 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p> <p>VALPROIC ACID 500 MG 24 HR (ER) SA TAB TAKE THREE TABLETS ORALLY AT BEDTIME</p> <p>TRAZODONE 50 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME AS NEEDED FOR INSOMNIA</p>
V2	<p>DISULFIRAM 250 MG TAB TAKE ONE TABLET ORALLY EVERY DAY</p> <p>VALPROIC ACID 500 MG 24 HR (ER) SA TAB TAKE THREE TABLETS ORALLY AT BEDTIME</p> <p>TRAZODONE 50 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME AS NEEDED FOR INSOMNIA</p>
PHCHP136V1	<p>BENZTROPINE MESYLATE ORAL MESYLATE 1 MG TAB TAKE ONE TABLET ORALLY TWICE A DAY</p> <p>CHLORPROMAZINE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>HALOPERIDOL DECANOATE 5 ML(100 MG/ML) INJ INJECT 200 MG HOLD(2 ML) INTRAMUSCULAR EVERY 4 WEEKS</p> <p>OXCARBAZEPINE 300 MG TAB TAKE ONE TABLET ORALLY EVERY MORNING AND TAKE THREE TABLETS AT BEDTIME</p> <p>FISH OIL CAP/TAB</p>
v2	<p>BENZTROPINE MESYLATE ORAL MESYLATE 2 MG TAB TAKE ONE TABLET ORALLY TWICE A DAY</p> <p>CHLORPROMAZINE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>HALOPERIDOL DECANOATE 5 ML(100 MG/ML) NJ INJECT 200 MG HOLD (2 ML) INTRAMUSCULAR EVERY 4 WEEKS</p> <p>OXCARBAZEPINE 300 MG TAB TAKE ONE TABLET ORALLY EVERY MORNING AND TAKE THREE TABLETS AT BEDTIME</p>
v3	<p>BENZTROPINE MESYLATE ORAL MESYLATE 2 MG TAB TAKE ONE TABLET ORALLY TWICE A DAY</p> <p>CHLORPROMAZINE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>HALOPERIDOL DECANOATE 5 ML(100 MG/ML) INJ INJECT 200 MG HOLD(2 ML) INTRAMUSCULAR EVERY 4 WEEKS</p> <p>OXCARBAZEPINE 300 MG TAB TAKE ONE TABLET ORALLY EVERY MORNING AND TAKE THREE TABLETS AT BEDTIME</p>
Phchp153v1	<p>TRAZODONE 50 MG TAB TAKE ONE TO ONE AND ONE-HALF TABLETS ORALLY AT BEDTIME</p> <p>VENLAFAXINE HCL 225 MG 24 HR SA TAB TAKE ONE TABLET ORALLY EVERY DAY WITH BREAKFAST</p>
v2	<p>TRAZODONE 100 MG TAB TAKE ONE TABLET ORALLY AT BEDTIME</p> <p>VENLAFAXINE HCL 225 MG 24 HR SA TAB TAKE ONE TABLET ORALLY EVERY DAY WITH BREAKFAST</p>
v3	<p>VENLAFAXINE HCL 150 MG 24 HR SA TAB-1X PER DAY</p> <p>TRAZADONE HCL 50 MG-1X PER DAY</p>

TABLE 2A-continued

Psychiatric medications of Discovery Cohort subjects.	
SubjectID-Visit	Psychiatric Medications
Phchp179v1	LISDEXAMFETAMINE (40 MG) QUETIAPINE (600 MG) PAROXETINE (30 MG) ALPRAZOLAM (½ MG PER NIGHT) ZOLPIDEM (10 MG PER NIGHT)
v2	No Psychiatric Medication
v3	QUETIAPINE 100 MG-IS BEING TAPERED OFF ZIPRASIDONE 120 MG PAROXETINE 30 MG ALPRAZOLAM unknown dosage, PRN LISDEXAMFETAMINE 50 MG ARIPIPRAZOLE TAB 20 MG PO DAILY PHCHP183V1 BENZTROPINE MESYLATE ORAL TAB 1 MG PO Q4H PRN VALPROIC ACID TAB, SA, 24 HR (EXPENDED) 2000 MG PO BEDTIME HALOPERIDOL INJ, SOLN 5 MG IM Q4H PRN HALOPERIDOL TAB 5 MG PO Q4H PRN HYDROXYZINE PAMOATE CAP, ORAL 25 MG PO Q6H PRN LORAZEPAM INJ 2 MG/1 ML IM Q4H PRN RISPERIDONE TAB 1 MG PO BID HYDROXYZINE PAMOATE 25 MG CAP TAKE ONE CAPSULE ORALLY EVERY 6 HOURS AS NEEDED FOR ANXIETY RISPERIDONE 1 MG TAB TAKE ONE-HALF TABLET ORALLY TWICE A DAY FISH OIL CAP/TAB ORALLY v2 ARIPIPRAZOLE 20 MG TAB TAKE ONE TABLET ORALLY EVERY DAY HYDROXYZINE PAMOATE 25 MG CAP TAKE ONE CAPSULE ORALLY EVERY 6 HOURS AS NEEDED FOR ANXIETY CITALOPRAM HYDROBROMIDE 10 MG TAB TAKE ONE-HALF TABLET ORALLY EVERY MORNING

TABLE 2B

Toxicology for subject in the coroner's office test cohort-suicide completers	
SubjectID	Toxicology
INBR009	
INBR011	ALPRAZOLAM 3.2 NG/ML TRAMADOL 331 NG/ML NORTRAMADOL 179 NG/ML BUPROPION 136 NG/ML CITALOPRAM/ESCITALOPRAM 229 NG/ML CAFFEINE POSITIVE COTININE POSITIVE
INBR012	Not Available
INBR013	CAFFEINE POSITIVE
INBR014	ETHANOL 0.15% (W/V) CAFFEINE
INBR015	ETHANOL 0.119% (W/V) CAFFEINE
INBR016	Not Available
INBR017	Not Available
INBR018	ETHANOL 0.057% (W/V) AMIODARONE CAFFEINE COTININE

[0055] The subjects were subjected to diagnostic assessments using Diagnostic Interview for Genetic Studies, which is the scale used by the Genetics Initiative Consortia for both bipolar disorder and major depression, at a baseline visit, followed by up to three testing visits, three to six months apart. Particularly, six subjects were subjected to three fol-

low-up testing visits and three subjects were subjected to two follow-up testing visits, resulting in a total of 24 blood samples for subsequent microarray studies as discussed herein. At each testing visit, the subjects received a series of psychiatric rating scales, including the Hamilton Rating Scale for Depression (HAMD-17), which includes a suicidal ideation rating item (FIG. 1B), and blood was drawn. The suicidal ideation scores varied during the visits from no ideation to high suicidal ideation.

Gene Expression Analysis

[0056] Using the nine subjects with multiple visits, corresponding to 24 chips, from the Discovery Cohort a differential analysis was run using Partek Genomic Suites 6.6 software package (Partek Incorporated, St. Louis, Mo.). Normalization was performed on all 24 chips by robust multi-array analysis (RMA), background corrected with quartile normalization and a median polish probe set summarization of all 24 chips to obtain the normalized expression levels of all probe sets for each chip. Two analyses, an intra-subject analysis and an inter-subject analysis, were conducted to establish a list of differentially expressed probe sets.

[0057] RNA Extraction.

[0058] During each visit, from about 2.5 ml to about 5.0 ml of whole blood was collected from the subjects separately into two PaxGene tubes, treated to stabilize RNA, by routine venipuncture. The cells from the whole blood were concentrated by centrifugation, the pellet washed, resuspended and incubated in buffers containing proteinase K for protein digestion. A second centrifugation step was conducted to remove residual cell debris. Ethanol was added. After ethanol addition, the supernatant was applied to a silica-gel membrane/column. The column was centrifuged and contaminants were removed in three wash steps. RNA bound to the membrane was then eluted using DEPC-treated water.

[0059] Globin Reduction.

[0060] To remove globin mRNA, total RNA from the whole blood was mixed with a biotinylated Capture Oligo Mix that is specific for human globin mRNA. The mixture was then incubated for 15 minutes to allow the biotinylated oligonucleotides to hybridize with the globin mRNA. Streptavidin magnetic beads were then added, and the mixture was incubated for 30 minutes. During this incubation, streptavidin binds to the biotinylated oligonucleotides, thereby capturing the globin mRNA on the magnetic beads. The streptavidin magnetic beads were then pulled to the side of the tube with a magnet, and the RNA, depleted of the globin mRNA, was transferred to a fresh tube. The treated RNA was further purified using a rapid magnetic bead-based purification method consisting of adding an RNA binding bead suspension to the samples and using magnetic capture to wash and elute the globin-clear RNA.

[0061] Sample Labeling.

[0062] Samples were labeled using an Ambion MessageAmp II-BiotinEnhanced antisense RNA (aRNA) amplification kit. The procedure involved the following steps:

[0063] 1) Reverse transcription to synthesize first strand cDNA was primed with T7 Oligo(dT) primer to synthesize cDNA containing a T7 promoter sequence.

[0064] 2) Second strand cDNA synthesis converted the single-stranded cDNA into a double-stranded DNA (ds-DNA) template for transcription. The reaction employed DNA polymerase and RNase H to simultaneously degrade the RNA and synthesize second strand cDNA.

[0065] 3) cDNA purification removed RNA, primers, enzymes, and salts that would inhibit in vitro transcription.

[0066] 4) In vitro transcription to synthesize aRNA with biotin-NTP mix generated multiple copies of biotin-modified aRNA from the double-stranded cDNA templates; this was the amplification step.

[0067] 5) aRNA purification removed unincorporated NTPs, salts, enzymes, and inorganic phosphate to improve the stability of the biotin-modified aRNA.

[0068] 6) aRNA fragmentation in a reaction that employs a metal-induced hydrolysis. The fragmented labeled aRNA is then ready for hybridization to the Affymetrix microarray chip.

[0069] Microarrays.

[0070] Biotin-labeled aRNA was then hybridized to Affymetrix HG-U133 Plus 2.0 GeneChips (Affymetrix, Santa Clara, Calif.) with over 40,000 genes and expressed sequence tags (ESTs) according to manufacturer's protocols (http://www.affymetrix.com/support/technical/manual/expression_manual.affx). All GAPDH 3'/5' ratios should be less than 2.0 and backgrounds under 50. Arrays were stained using standard Affymetrix protocols for antibody signal amplification and scanned on an Affymetrix GeneArray 2500 scanner with a target intensity set at 250. Present/absent calls were determined using GCOS software with thresholds set at default values. Quality control measures including 30/50 ratios for glyceraldehyde 3-phosphate dehydrogenase and b-actin, scale factors, background and Q values were within acceptable limits.

[0071] Analysis.

[0072] Each subject's suicidal ideation (SI) scores at time of blood collection (0—no SI compared to 2 and above—high SI) were used for analysis. Particularly, gene expression differences between the no SI and the high SI states using both an intra-subject and an inter-subject design as shown in FIG. 1A were analyzed.

[0073] An intra-subject analysis using a fold change in expression of at least 1.2 between high and no suicidal ideation visits within each subject was performed. There were in total 15 comparisons. Probe sets that had a 1.2 fold change were then assigned either a 1 (increased in high suicidal ideation) or a -1 (decreased in high suicidal ideation) in each comparison. These values were then summed for each probe set across the 15 comparisons, yielding a range of scores between -11 and 12. Probe sets in the top 5% (1,269 probe sets, <5% of 54,675 total probe sets) had an absolute value of 7 and greater, receiving an internal Convergent Functional Genomics (CFG) score of 1 point. Those probe sets in the top 0.1% (24 probe sets, <0.1% of 54,675 total probe sets) had a total absolute value of 11 and greater and received an internal CFG score of 3 points.

[0074] Additionally, an inter-subject analysis using t-test (2-tailed, unequal variance) was performed to find probes differentially expressed between high suicidal ideation and no suicidal ideation chips (FIG. 1A), resulting in 648 probe sets with $P < 0.05$. Probe sets with a $P < 0.05$ received an internal CFG score of 1 point, while probe sets with $P < 0.001$ received 3 points.

[0075] Results were then further filtered by only selecting probe sets that overlapped between the intra-subject and the inter-subject analyses, resulting in 279 probe sets corresponding to 246 unique genes. Gene names for the probe sets were identified using Partek as well as NetAffy (Affymetrix) for

Affymetrix HG-U133 Plus 2.0 GeneChips, followed by GeneCards to confirm the primary gene symbol. In addition, for those probe sets that were not assigned a gene name by Partek or NetAffy, the UCSC Genome Browser on Human February 2009 (GRCh37/hg19) was used to directly map them to known genes. Genes were then scored using manually curated CFG databases as described below and shown in FIG. 2.

Convergent Functional Genomics (CFG) Databases

[0076] Manually curated databases were created in the Laboratory of Neurophenomics, Indiana University School of Medicine (www.neurophenomics.info) of all the human gene expression (postmortem brain, blood, cell cultures), human genetic (association, CNVs, linkage) and animal model gene expression and genetic studies published to date on psychiatric disorders. Only the findings deemed significant in the primary publication, by the study authors, using their particular experimental design and thresholds, were included in the databases. The databases included only primary literature data and did not include review papers or other secondary data integration analyses to avoid redundancy and circularity. These large and constantly updated databases have been used in previous CFG cross-validation and prioritization studies.

[0077] Human Postmortem Brain Gene Expression Evidence.

[0078] Information about genes was obtained and imported in the databases searching the primary literature with PubMed (<http://ncbi.nlm.nih.gov/PubMed>), using various combinations of keywords (e.g., gene name and suicide and human brain). Postmortem convergence was deemed to occur for a gene if there were published reports of human postmortem data showing changes in expression of that gene in brains from patients who died from suicide.

[0079] Human Genetic Evidence Association and Linkage.

[0080] To designate convergence for a particular gene, the gene had to have independent published evidence of association or linkage for suicide. For linkage, the location of each gene was obtained through GeneCards (<http://www.genecards.org>), and the sex averaged cM location of the start of the gene was then obtained through <http://compngen.rutgers.edu/old/map-interpolator/>. For linkage convergence, the start of the gene had to map within 5 cM of the location of a marker linked to the disorder.

[0081] CFG Scoring.

[0082] For CFG analysis, two external cross-validating lines of evidence were weighed such that findings in human postmortem brain tissue, the target organ, were prioritized over genetic findings. Human brain expression evidence was given 4 points, while human genetic evidence was given a maximum of 2 points for association, and 1 point for linkage. Each line of evidence was capped in such a way that any positive findings within that line of evidence resulted in maximum points regardless of how many different studies support that single line of evidence, to avoid potential popularity biases.

[0083] In addition to the above external CFG score, genes based upon the initial differential expression analyses used to identify them were also prioritized. Probe sets identified by differential expression analyses could receive a maximum of 6 points (1 or 3 points from intra-subject analyses, and 1 or 3 points from inter-subject analyses). Thus, the maximum possible total CFG score for each gene was 12 points (6 points for internal score+6 points for external score).

[0084] The above-described scoring system provided a good separation of genes based on differential expression and on independent cross-validating evidence in the field (FIG. 2).

Pathway Analyses

[0085] IPA 9.0 (Ingenuity Systems, www.ingenuity.com, Redwood City, Calif.) was used to analyze the biological roles, including top canonical pathways and diseases, of the candidate genes resulting from the above findings (Table 3), as well as used to identify genes in the data sets that were the target of existing drugs (Table 4). Pathways were identified

from the IPA library of canonical pathways that were most significantly associated with genes in the data set. The significance of the association between the data set and the canonical pathway was measured in two ways: 1) a ratio of the number of molecules from the data set that map to the pathway divided by the total number of molecules that map to the canonical pathway is displayed; and 2) Fisher's exact test was used to calculate a p-value determining the probability that the association between the genes in the data set and the canonical pathway was explained by chance alone. A KEGG pathway analysis through the Partek Genomic Suites 6.6 software package was also conducted.

TABLE 3

Ingenuity Pathway Analyses.							
A. Pathways.							
B. Disease and Disorders.							
A.							
INGENUITY Pathways			KEGG Pathways				
	Top Canonical # Pathways	P-Value	Ratio	Pathway Name	Enrichment Score	Enrichment p-value	
CFG score \geq 6.0 N = 21 genes	1	Role of Tissue Factor in Cancer	2.63E-04	3/115 (0.026)	Apoptosis	6.69102	0.001242
	2	Dendritic Cell Maturation	9.83E-04	3/207 (0.014)	Measles	6.06369	0.002326
	3	Melanoma Signaling	1.13E-03	2/46 (0.043)	Endometrial cancer	4.96787	0.006958
	4	Docosahexaenoic Acid (DHA) Signaling	1.18E-03	2/49 (0.041)	Influenza A	4.90223	0.00743
	5	Endometrial Cancer Signaling	1.69E-03	2/57 (0.035)	Phosphatidylinositol signaling system	4.85448	0.007793
CFG score \geq 4.0 N = 41 genes	1	NF- κ B Signaling	4.42E-04	4/175 (0.023)	Measles	8.7667	0.000156
	2	Dendritic Cell Maturation	5.38E-04	4/207 (0.019)	Influenza A	6.87308	0.001035
	3	PDGF Signaling	7.5E-04	3/85 (0.035)	mTOR signaling pathway	6.34986	0.001747
	4	Role of Pattern Recognition Receptors in Recognition of Bacteria and Viruses	1.14E-03	3/106 (0.028)	Apoptosis	4.75687	0.008592
	5	Role of Tissue Factor in Cancer	1.78E-03	3/115 (0.026)	Toll-like receptor signaling pathway	4.37269	0.012617
All genes differentially expressed N = 246 genes (279 probe sets)	1	Retinoic acid Mediated Apoptosis Signaling	1.12E-03	5/69 (0.072)	Ubiquitin mediated proteolysis	4.80416	0.0081956
	2	Role of PKR in Interferon Induction and Antiviral Response	1.19E-03	4/46 (0.087)	Herpes simplex infection	4.14288	0.0158771
	3	UVA-induced MAPK Signaling	3.90E-03	5/92 (0.054)	Phagosome	4.0301	0.0177725
	4	Dendritic Cell Maturation	4.71E-03	7/207 (0.034)	Measles	3.72158	0.0241958
	5	Role of Pattern Recognition Receptors in Recognition of Bacteria and Viruses	5.38E-03	5/106 (0.047)	Influenza A	5.03358	0.0065155

TABLE 3-continued

Ingenuity Pathway Analyses.				
A. Pathways.				
B. Disease and Disorders.				
B.				
INGENUITY				
	#	Diseases and Disorders	P-Value	# Molecules
CFG score \geq 6.0 N = 21 genes	1	Cancer	1.22E-06-4.54E-03	14
	2	Connective Tissue Disorders	2.19E-04-3.41E-03	8
	3	Inflammatory Disease	2.19E-04-4.54E-03	8
	4	Skeletal and Muscular Disorders	2.19E-04-4.42E-03	9
	5	Gastrointestinal Disease	2.22E-04-4.54E-03	12
CFG score \geq 4.0 N = 41 genes	1	Cancer	4.51E-06-6.45E-03	20
	2	Inflammatory Response	2.70E-05-6.45E-03	12
	3	Antimicrobial Response	9.95E-05-6.45E-03	4
	4	Infectious Disease	1.25E-04-5.52E-03	6
	5	Connective Tissue Disorders	1.53E-04-6.45E-03	11

TABLE 4

Ingenuity drug targets analysis. Repositioning of existing drugs for treating suicidality.					
	CFG Score	Direction of change	Location	Type(s)	Drug(s)
IL1B interleukin 1, beta	8	I	Extracellular Space	cytokine	IL-1 trap, canakinumab
KCNJ2 potassium inwardly-rectifying channel, subfamily J, member 2	4	I	Plasma Membrane	ion channel	nicorandil, amiodarone
PML promyelocytic leukemia	4	I	Nucleus	transcription regulator	arsenic trioxide
TNK2 tyrosine kinase, non-receptor, 2	4	D	Cytoplasm	kinase	vemurafenib
TOP1 topoisomerase (DNA) I	2	I	Nucleus	enzyme	elsamitucin, T 0128,CT-2106, BN 80927, tafluposide, TAS- 103, beta- lapachone, irinotecan, topotecan, 9- amino-20- camptothecin, rubitecan, gimatecan, karenitecin

Validation Analyses

[0086] The nine Affymetrix microarray data files from the Validation Cohort was imported as .cel files into Partek Genomic Suites 6.6 software package (Partek Incorporated St. Louis, Mo.). A robust multi-array analysis (RMA), background corrected with quartile normalization and a median polish probe set summarization of all 24+9=33 chips was conducted to obtain the normalized expression levels of all probe sets for each chip. Partek normalizes expression data into a log base of 2 for visualization. The data was non-log by taking 2 to the power of the transformed expression value. The non-log transformed expression data was then used to compare expression levels of SAT1 and CD24 in the different groups (FIG. 3G).

[0087] Further, testing of the top candidate biomarkers for suicidality were conducted (see FIGS. 3H and 3I). As shown, thirteen of the 41 CFG-top scoring biomarkers from Table 5 below showed step-wise significant change from no suicide ideation to high suicide ideation, to the validation suicide completers group. Six of them (15%) remained significant after strict Bonferroni correction for multiple comparisons. The top CFG scoring biomarker, SAT1, remained the top biomarker after validation.

Results

[0088] Whole-genome gene expression profiling in blood samples from a longitudinally-followed homogeneous cohort of male subjects with a major mood disorder (bipolar disorder)

der) that predisposes to suicidality was conducted. The samples were collected at repeated visits, 3 to 6 months apart. State information about suicidal ideation was collected from a questionnaire administered at the time of each blood draw. An intra-subject design was used to analyze data from 9 subjects that switched from no suicidal ideation to high suicidal ideation at different visits, which factors out genetic variability, as well as some medications, lifestyle and demographic variability. An inter-subject case-case analysis was also used to identify genes differentially expressed in the blood in no suicidal ideation states versus high suicidal ideation states. The top 0.1% and 5% of the gene expression probe sets distributions were considered and differentially scored. Overlap between the intra-subject and inter-subject analyses of gene expression changes was required. Such a restrictive approach was used as a way of minimizing false positives, even at the risk of having false negatives. For example, there were genes on each of the two lists, from intra- and inter-subject analyses, that had clear prior evidence for involvement in suicidality, such as MT1E (Sequeira A. et al., Gene expression changes in the prefrontal cortex, anterior cingulate cortex and nucleus accumbens of mood disorders subjects that committed suicide, *PloS one* 7, e35367, doi:10.1371/journal.pone.0035367 (2012)), respectively GSK3B (Karege F. et al., Alteration in kinase activity but not in protein levels of protein kinase B and glycogen synthase kinase-3beta in ventral prefrontal cortex of depressed suicide victims. *Biol Psychiatry* 61, 240-245, doi:10.1016/j.biopsych.2006.04.036

(2007)), but were not included in the subsequent analyses because they were not in the overlap.

[0089] A CFG approach was then used to cross-match the list of 246 overlapping top differentially expressed genes from the blood samples with other key lines of evidence (human postmortem brain data, human genetic data) implicating them in suicidality, as a way of identifying and prioritizing disease-relevant genomic biomarkers, extracting generalizable signal out of potential cohort-specific residual noise and genetic heterogeneity. Manually curated databases of the psychiatric genomic and proteomic literature to date was created and used in the CFG analyses. The CFG approach was thus a de facto field-wide collaboration. In essence, in a Bayesian fashion, the whole body of knowledge in the field was used to leverage findings from the Discovery Cohort data sets. Unlike the use of CFG in previous studies, no human peripheral tissue evidence from the literature was used as there was none directly matching the instant genes, reflecting perhaps the dearth of peripheral gene expression work done so far on suicides, and the need for a study like the instant Example. Animal model evidence was also not used as there were to date no clear studies in animal models of self-harm or suicidality. SAT1 (spermidine/spermine N1-acetyltransferase 1) was the top blood biomarker increased in suicidal states (i.e. the top risk marker), and CD24 (CD24 molecule; small cell lung carcinoma cluster 4 antigen) was the top blood biomarker decreased in suicidal states (i.e. the top protective marker) (FIG. 2 and Table 5).

TABLE 5

Top gene expression biomarkers for suicidality						
Gene Symbol/ Gene name	Probe-sets	Change (I = Increase) (D = Decrease)	Differential Expression Score	Prior Human Genetic Evidence	Prior Human Brain Expression Evidence	Total CFG Score
SAT1 spermidine/spermine N1- acetyltransferase 1	203455_s_at	I	2	(Assoc) Suicide attempt (Fiori, Wanner et al. 2010). Suicide (Sequeira, Gwadry et al. 2006)	Suicide in Depression (D) PFC (Chen. Fiori et al., 2010) Suicide(D) AMY, PFC, HIP, THAL (Fiori, Bureau et al. 2011) Suicide(D) PFC (Flori and Turacki 2011) Suicide (D) PFC (Fiori, Mechawar et al. 2009) Suicide (D) PFC (Fiori, Zouk et al. 2011) Suicide(D) PFC (Guipponi, Deutsch et al. 2009) Suicide(D)PFC (Klempen, T. A. et al 2009) Suicide(D)PFC (Sequeira, A. et al. 2006)	8

TABLE 5-continued

Top gene expression biomarkers for suicidality						
Gene Symbol/ Gene name	Probe-sets	Change (I = Increase) (D = Decrease)	Differential Expression Score	Prior Human Genetic Evidence	Prior Human Brain Expression Evidence	Total CFG Score
CD24 CD24 molecule	209772_s_at	D	4		Suicide in mood disorder(D)NAC (Sequeira A. et al. 2012)	8
FOXP3 forkhead box N3	230790_x_at	I	2	(Assoc) Suicide (Galfalvy, H. et al. 2011)	Suicide (I) PFC (Galfalvy, H. et al. 2011)	8
GBP1 guanylate binding protein 1, interferon-inducible, 67 kDa	231577_x_at 202269_x_at 202270_at	I	4 2 2		Suicide in mood disorders (D) NAC (Karege, F. et al. 2007).	8 6 6
PIK3R5 Phosphoinositide-3- kinase, regulatory subunit5	227553_at	I	4		Suicide in mood disorder (D) PFC (Sequeira, Morgan et al. 2012)	8
APOL2 Apolipoprotein L2	221653_x_at	I	2		Suicide PFC (I) (Kekesi, K. A. et al. 2012)	6
ATP13A2 ATPase type 13A2	218608_at	D	2		Suicide(D) (Sequeira, A. et al. 2012)	6
ATP6V0E1 ATPase, H+ transporting, lysosomal 9 kDa, V0 subunit e1	214149_s_at 214244_s_at	I	2		Suicide(D)PFC (Sequeira A. et al. 2006)	6
EPHX1 epoxide hydrolase 1, microsomal (xenobiotic)	202017_at	D	2		Suicide in Schizophrenia (D) PFC (Kim, Choi et al. 2007)	6
GCOM1 GRINL1A complex locus	239099_at	I	2		Suicide in Depression (D) Klempman T A, 2009	6
HTRA1 HtrA serine peptidase 1	201185_at	D	2		Suicide(I) (Sequeira, A. et al. 2012)	6
IL1B interleukin 1, beta	39402_at	I	2		Suicide(1) PFC (Pandey, G. N. et al., 2012)	6
LEPR leptin receptor	211354_s_at	D	2		Suicide(D) PFC (Klempman, T. A. et al. 2009) Suicide(D) PFC (Lalovic, Klempen et al. 2010) Suicide(D) HIP (Sequeira, A. et al. 2007) Suicide in Depression (I) PFC (Zhurov V. et al. 2012)	6
LHFP lipoma HMGIC fusion partner	218656_s_at	I	2		Suicide in mood disorder (I) NAC (Sequeira, A. et al. 2012)	6
LIPA lipase A	236156_at	I	2		Violent Suicide (I) PFC (Freemantle, E et al. 2013)	6
MARCKS myristoylated alanine- rich protein kinase C substrate	213002_at	I	2		Suicide in Depression (I) (Pandey, G. N. et al. 2003)	6
PGLS 6-Phosphogluconolactonase	230699_at	I	2		Suicide PFC (D) (Kekesi K. A. et al. 2012)	6

TABLE 5-continued

Top gene expression biomarkers for suicidality						
Gene Symbol/ Gene name	Probe-sets	Change (I = Increase) (D = Decrease)	Differential Expression Score	Prior Human Genetic Evidence	Prior Human Brain Expression Evidence	Total CFG Score
PTEN phosphatase and tensin homolog	222176_at	I	2		Suicide PFC, HIP (I) (Dwivedi Y. et al. 2010)	6
RECK reversion-inducing- cysteine-rich protein with kazal motifs	216153_x_at	I	2		Suicide (I) PFC (Sequeira A. et al. 2012)	6
SPTBN1 spectrin, beta, non- erythrocytic 1	200671_s_at	D	2		Suicide in mood disorders (I) NAC (Sequeira A. et al. 2012)	6
TNFSF10 tumor necrosis factor (ligand) superfamily, member 10	202688_at 202687_s_at 214329_x_at	I	2		Suicide in Schizophrenia (I)PFC (Kim, S. et al. 2007) Suicide in Depression (I) PFC (Zhurov V. et al. 2012)	6
ABCA1 ATP-binding cassette, sub-family A (ABC1), member 1	203504_s_at	I	4			4
ARHGEF40 (FLJ10357) Rho guanine nucleotide exchange factor (GEF) 40	241631_at	I	4			4
CASC1 cancer susceptibility candidate 1	220168_at	I	4			4
DHRS9 dehydrogenase/reductase (SDR family) member 9	219799_s_at	I	4			4
DISC1 disrupted in schizophrenia 1	244642_at	I	2	(Assoc) Suicide (Galfalvy H. et al. 2011)		4
EIF2AK2 eukaryotic translation initiation factor 2-alpha kinase 2	204211_x_at	I	4			4
LOC727820 uncharacterized LOC727820	231247_s_at LOC727820	I	4			4
MAP3K3 mitogen-activated protein kinase kinase kinase 3	242117_at	I	4			4
MBNL2 muscleblind-like 2 (<i>Drosophila</i>)	205017_s_at	D	2	(Assoc) Suicide (Galfalvy H. et al. 2011)		4
MT-ND6 (ND6) mitochondrially encoded NADH dehydrogenase 6	1553575_at	I	4			4
OR2J3 olfactory receptor, family 2, subfamily J, member 3	217334_at	D	4			4
RBM47 RNA binding motif protein 47	1565597_at	I	4			4
RHEB Ras homolog enriched in brain	227633_at	D	2	(Assoc) Suicide (Menke A. et al. 2012)		4

TABLE 5-continued

Top gene expression biomarkers for suicidality						
Gene Symbol/ Gene name	Probe-sets	Change (I = Increase) (D = Decrease)	Differential Expression Score	Prior Human Genetic Evidence	Prior Human Brain Expression Evidence	Total CFG Score
RICTOR RPTOR independent companion of MTOR, complex 2	228248_at	I	4			4
SAMD9L sterile alpha motif domain containing 9-like	243271_at; 230036_at	I	4			4

[0090] In order to validate the Discovery Cohort findings in the most stringent way possible, SAT1 levels in blood samples from the Validation Cohort of 9 consecutive male suicide completers obtained from the coroner’s office were evaluated. SAT1 gene expression levels were found to be elevated in 9 out of 9 (100%) subjects who committed suicide. In each suicide completer, the increase in SAT1 was at least three standard deviations above the average levels in high suicidal ideation subjects. The results were further strengthened by using a panel of the two markers (SAT1 and CD24) (FIGS. 3A-C). As shown in FIGS. 3A-3C, risk marker SAT1 expression was significantly increased (p=0.0057) between subjects with high suicidal ideation (SI) (mean=3413.37) and those reporting no suicidal ideation (mean=2642.97). In the Validation Cohort of suicide completers (mean=7171.51), a significantly greater expression of SAT1 was found as compared to both high suicide ideation (p=7.27e-07) and no suicide ideation (p=1.51e-07) groups from the Discovery Cohort (FIG. 3A). Further, suicide risk score was calculated by scoring the standard deviation band a subject fell within as derived from the high suicide ideation Discovery Cohort, starting from the mean of the high suicide ideation Discovery Cohort (FIG. 3B). 0 indicates the subject fell between the means of the high and low suicide ideation subjects in the Discovery Cohort. A score of 1 means between the mean of the high suicide ideation and the first standard deviation above it, score of 2 between the first and second standard deviation, score of 3 between the second and third standard deviation, and so on.

[0091] As shown in FIGS. 3D-3F, protective marker CD 24 expression was significantly decreased (p=0.0044) within the Discovery Cohort between subjects reporting high suicide ideation (mean=73.01) and no suicide ideation (mean=108.634). The Validation Cohort of suicide completers (mean=71.61) was also significantly decreased (p=0.0031) when compared to subjects reporting no suicide ideation (FIG. 3D). Suicide risk score was defined as the standard deviation band in which the subject expression fell below the mean of the high suicide ideation Discovery Cohort (FIG. 3E).

[0092] FIG. 3G shows the sum of standard deviation suicide risk scores for both biomarkers (SAT1 and CD24) in the Validation Cohort (i.e., suicide completers).

[0093] One of the other biomarkers identified to be decreased in high suicidal states in the current Example was the circadian clock gene DBP (D-box binding protein). Serendipitously, previous work showed that mice engineered to lack DBP were stress-reactive and displayed a behavioral phenotype similar to bipolar disorder and co-morbid alcoholism (Le-Niculescu H. et al., “Phenomic, convergent functional genomic, and biomarker studies in a stress-reactive genetic animal model of bipolar disorder and co-morbid alcoholism,” American Journal of Medical Genetics, Part B, Neuropsychiatric genetics: the official publication of the International Society of Psychiatric Genetics 147B, 134-166, doi:10.1002/ajmg.b.30707 (2008)). In addition to bipolar disorder, alcoholism is known to increase the risk for suicide. Treatment with omega-3 fatty acids normalized the phenotype of those mice. Low omega-3 levels have been previously correlated with increased suicidality in human epidemiological studies (see Sublette M. et al., “Omega-3 polyunsaturated essential fatty acid status as a predictor of future suicide risk,” Am J Psychiatry 163, 1100-1102, doi:10.1176/appi.ajp.163.6.110 (2006); Lewis M. D. et al., “Suicide deaths of active-duty US military and omega-3 fatty-acid status: a case-control comparison,” J Clin Psychiatry 72, 1585-1590, doi:10.4088/JCP.11m06879 (2011)). Pathway analyses of the instant suicidality biomarker data identified among the top pathways the omega-3 docosahexaenoic acid (DHA) signaling pathway. Several of the biomarkers from this Example (those bolded in Table 6 in “Modulated by DHA” column)) were changed in expression by omega-3 treatment in the blood of the DBP mouse model in opposite direction to our human suicidality data (Table 6). PTEN, a biomarker increased in suicidality in the current Example in the blood, as well as in the brain of suicide completers, was also increased in the amygdala and decreased in the pre-frontal cortex of DBP knock-out mice subjected to stress.

TABLE 6

Genes in our dataset modulated by Clozapine and Omega-3 Fatty Acids (DHA).					
Gene Symbol	Gene Name	Direction of Change	CFG score	Modulated by Clozapine	Modulated by DHA
SAT1	spermidine/spermine N1-acetyl transferase 1	I	8		(D) Blood
GBP1	guanylate nucleotide binding protein 1	I	8		(D) Blood
ATP13A2	ATPase type 13A2	D	6	(D) VT	
EPHX1	epoxide hydrolase 1, microsomal	D	6	(D) VT	

TABLE 6-continued

Genes in our dataset modulated by Clozapine and Omega-3 Fatty Acids (DHA).					
Gene Symbol	Gene Name	Direction of Change	CFG score	Modulated by Clozapine	Modulated by DHA
IL1B	interleukin 1 beta	I	6	(I) Blood	(D) Blood
LHFP	lipoma HMGIC fusion partner	I	6	(I) Blood, VT	(D) Blood
MARCKS	myristoylated alanine rich protein kinase C substrate	I	6		(I) HIP
PTEN	phosphatase and tensin homolog	I	6	(I) VT	
SPTBN1	spectrin, beta, non-erythrocytic 1	D	6	(I) Blood, VT	(D) Blood
ABCA1	ATP-binding cassette, sub-family A (ABC1), member 1	I	4	(I) VT	
MAP3K3	mitogen-activated protein kinase kinase kinase 3	I	4		(D) Blood
MBNL2	muscleblind-like 2	D	4	(I) Blood	(D) Blood
ATG3	autophagy-related 3 (yeast)	I	2		(D) Blood
ATXN2	ataxin 2	I	2	(I) VT	
CCR1	chemokine (C-C motif) receptor 1	I	2		(D) Blood
CCRN4L	CCR4 carbon catabolite repression 4-like	I	2		(I) Blood
CD84	CD84 antigen	D	2	(I) Blood	(D) Blood
CEACAM1	CEA-related cell adhesion molecule 1	I	2		(D) Blood
CELA1	chymotrypsin-like elastase family, member 1	D	2		(D) Blood
CLEC4E	C-type lectin domain family 4, member e	I	2		(D) Blood
CLEC7A	C-type lectin domain family 7, member a	I	2		(D) Blood
CORO1C	coronin, actin binding protein 1C	I	2	(D) VT	
DLGAP1	discs, large (<i>Drosophila</i>) homolog-associated protein 1	I	2	(I) VT	
DOCK1	dedicator of cytokinesis 1	D	2	(D) VT	
DOCK4	dedicator of cytokinesis 4	I	2	(D) HIP	
FABP3	fatty acid binding protein 3, muscle and heart	I	2	(I) VT	
FNIP1	folliculin interacting protein 1	I	2	(I) VT	
FOXK2	forkhead box K2	D	2	(I) VT	
FZR1	fizzy/cell division cycle 20 related 1 (<i>Drosophila</i>)	D	2		(I) Blood
GBP2	guanylate nucleotide binding protein 2	I	2	(D) VT	
GREM1	gremlin 1	I	2		(D) HIP
IFIT2	interferon-induced protein with tetratricopeptide repeats 2	I	2	(I) Blood	(D) Blood
IFIT3	interferon-induced protein with tetratricopeptide repeats 3	I	2		(D) NAC
IL1RAP	interleukin 1 receptor accessory protein	I	2	(I) VT	
KLHDC3	kelch domain containing 3	D	2	(I) VT	
KPNA3	karyopherin (importin) alpha 3	I	2	(D) VT	
LARP4	La ribonucleoprotein domain family, member 4	D	2	(I) VT	
LONRF1	LON peptidase N-terminal domain and ring finger 1	I	2	(I) VT	
MCTP1	multiple C2 domains, transmembrane 1	I	2		(I) HIP
MDM4	transformed mouse 3T3 cell double minute 4	I	2		(D) Blood
NUB1	negative regulator of ubiquitin-like proteins 1	I	2	(D) VT	
NUDT3	nudix (nucleotide diphosphate linked moiety X)-type motif 3	I	2		(D) Blood
OGT	O-linked N-acetylglucosamine (GlcNAc) transferase	I	2	(I) Blood	(D) HIP; (I) NAC
PELL1	pellino 1	I	2	(I) AMY	(I) HIP

TABLE 6-continued

Genes in our dataset modulated by Clozapine and Omega-3 Fatty Acids (DHA).					
Gene Symbol	Gene Name	Direction of Change	CFG score	Modulated by Clozapine	Modulated by DHA
PKN2	protein kinase N2	I	2		(I) Blood
R3HDM1	R3H domain 1 (binds single-stranded nucleic acids)	I	2	(I) VT	(D) Blood
RAI14	retinoic acid induced 14	D	2		(I) HIP
RASSF3	Ras association (RalGDS/AF-6) domain family member 3	I	2	(I) VT	(D) Blood
RPL37A	ribosomal protein L37a	I	2		(I) Blood
RPLP2	ribosomal protein, large P2	I	2		(I) Blood
RSAD2	radical S-adenosyl methionine domain containing 2	I	2	(I) Blood	(I) Blood
S100A8	S100 calcium binding protein A8 (calgranulin A)	I	2	(D) Blood	(D) Blood
SFRP2	secreted frizzled-related protein 2	D	2	(D) VT	
SLC25A37	solute carrier family 25, member 37	I	2	(I) VT	(I) Blood
SLC2A13	solute carrier family 2 (facilitated glucose transporter), member 13	I	2	(I) VT	
SPOCK2	sparc/osteonectin, cwcv and kazal-like domains proteoglycan 2	D	2	(I) VT	
TAOK1	TAO kinase 1	I	2	(I) VT	(D) PFC; (I) HIP
TB1X	transducin (beta)-like 1 X-linked	I	2	(D) VT	
TCEA1	transcription elongation factor A (SII) 1	I	2	(D) VT	(I) Blood
TMEM140	transmembrane protein 140	I	2	(I) Blood	(D) Blood
TMEM154	transmembrane protein 154	I	2	(I) Blood	(D) Blood
TNFAIP6	tumor necrosis factor alpha induced protein 6	I	2	(I) AMY	
TNK2	tyrosine kinase, non-receptor, 2	D	2	(D) VT	
TOP1	topoisomerase (DNA) I	I	2	(I) VT	(I) Blood
TRIP12	thyroid hormone receptor interactor 12	I	2	(I) VT	
TRPM7	transient receptor potential cation channel, subfamily M, member 7	I	2		(D) AMY
UBE2B	ubiquitin-conjugating enzyme E2B, RAD6 homology (S. cerevisiae)	I	2	(I) Blood, AMY, PFC	(I) Blood
WDR77	WD repeat domain 77	D	2	(D) VT	

Bold are genes that are changed in opposite direction to suicidal ideation by one or both of the treatments.

[0094] Other circadian clock-modulated genes identified as biomarkers for suicidality were PIK3R5, MARCKS, IL1B, CASC1, CCRN4L, H3F3B, RBCK1, TNK2, and UBE2B. Additionally, biomarkers, as bolded in Table 6 in the “Modulated by Clozapine” column, provided evidence for modulation by clozapine in blood in opposite direction to the human suicidality data in previous independent animal model pharmacogenomics studies (Table 6). Clozapine is the only FDA approved treatment for suicidality. Thus, the convergent evidence for the instant biomarkers is strong in translational ways beyond those used for their discovery and selection. S100A8 may be a key biomarker to monitor in terms of response to treatment with classic (clozapine) and complementary (omega-3) agents. Other potential drugs to be studied for modulating suicidality were revealed by the above analyses (Tables 4 and 6).

[0095] SAT1, FOXN3, DISC1, MBNL2 and RHEB had genetic association evidence for suicidality, suggesting that they are not only state biomarkers but also trait factors influ-

encing suicidal risk. DISC1 is also one of the top candidate genes for schizophrenia based on a large scale CFG analysis of schizophrenia GWAS recently conducted (Ayalew M. et al., “Convergent functional genomics of schizophrenia: from comprehensive understanding to genetic risk prediction,” *Molecular Psychiatry* 17, 887-905, doi:10.1038/mp.2012.37 (2012)), while DISC1 and MBNL2 are also among the top candidate genes for bipolar disorder based on a large scale CFG analysis of bipolar disorder GWAS (Patel S. D. et al., “Coming to grips with complex disorders: genetic risk prediction in bipolar disorder using panels of genes identified through convergence functional genomics,” *American Journal of Medical Genetics Part b, Neuropsychiatric genetics: the official publication of the International Society of Psychiatric genetics* 153B, 850-877, doi:10.1pp2/ajmg.b.31087 (2010)). Additionally, DISC1 has clear animal model data for the role of its interaction with environmental stress in the pathophysiology of psychotic depression. DISC1 and MBNL2 may thus be key state and trait factors for suicidality

risk in psychotic mood disorder subjects, and an indication for clozapine treatment in such subjects.

[0096] Suicide biomarkers that were identified in this study were overlapped with biomarkers identified as mood biomarkers (Le-Niculescu H. et al., “Identifying blood biomarkers for mood disorders using convergent functional genomics,” *Molecular Psychiatry* 14, 156-174, doi:10.1111/ele.12064 (2009)) and psychosis biomarkers (Kurian S. M. et al., “Identification of blood biomarkers for psychosis using convergent functional genomics,” *Molecular Psychiatry* 16, 37-58, doi:

10.1038/mp.2009.117 (2011)) (Table 7). DOCK5 and 4 other biomarkers (as bolded in Table 7 in the “Direction of change in Mood” column were changed in high suicidal states in opposite direction to their change in high mood states, and DOCK5 and 6 other biomarkers (as bolded in Table 7 in the “Direction of change in Hallucination” or “Direction of change in Delusions” columns) were changed in the same direction as their change in high psychosis states, suggesting that suicidality can indeed be viewed as a psychotic depressed state, and that DOCK5 may be an additional key biomarker reflecting that state.

TABLE 7

Genes with evidence as mood and/or psychosis blood biomarkers.						
Gene Symbol	Gene Name	CFG score in SI	Direction of change in SI	Direction of change in Mood	Direction of change in Hallucination	Direction of change in Delusions
LEPR	leptin receptor	6	D	(I)		
CD84	CD84 molecule	2	D			(I)
DOCK5	dedicator of cytokinesis 5	2	I	(D)	(I)	
EPM2A	epilepsy, progressive myoclonus type 2A, Lafora disease (laforin)	2	D			(D)
ERICH1	glutamate-rich 1	2	I		(D)	(D)
FKBP7	FK506 binding protein 7	2	D			(D)
IDH1	isocitrate dehydrogenase 1 (NADP+), soluble	2	I			(D)
KIAA0494	KIAA0494	2	I	(D)		
LARP4	La ribonucleoprotein domain family, member 4	2	D	(D)		
MXD1	MAX dimerization protein 1	2	I			(I)
PID1	phosphotyrosine interaction domain containing 1	2	D			(D)
PML	promyelocytic leukemia protein	2	I			(I)
PPP2R1B	phosphatase 2, regulatory subunit A, beta	2	D			(D)
SLC2A13	solute carrier family 2 (facilitated glucose transporter), member 13	2	I	(D)		
TRIM6	tripartite motif containing 6	2	I	(D)		
TRPM7	transient receptor potential cation channel, subfamily M, member 7	2	I	(I)		

DISCUSSION

[0097] This Example shows overlap at a gene and pathway level with cancer and apoptosis (Table 3, Table 8). SAT1, for example, is a key catabolic enzyme for polyamines. Polyamine levels within cells control cell viability, and sig-

nificant decreases in polyamine levels can result in apoptosis. They appear to reflect an endowment for cellular and organ- ismal activity and growth, key characteristics of mood. SAT1, which increased in suicidal subjects of this Example, is highly inducible by a variety of stimuli, including toxins, cytokines, heat shock, ischemia, and other stresses.

TABLE 8

Complete list of genes differentially expressed in the discovery cohort overlapping between the intra-subject and inter-subject analyses (n = 246).					
Probe set ID	Gene Symbol	Gene Name	Change	Total CFG Score	Evidence or possible roles in apoptosis
203455_s_at	SAT1	spermidine/spermine N1-acetyltransferase 1	I	8	yes
209772_s_at	CD24	CD24 molecule	D	8	yes
230790_x_at	FOXN3	forkhead box N3	I	8	
231577_s_at;	GBP1	guanylate binding protein 1,	I	8	yes
202269_x_at;		interferon-inducible			
202270_at					
227553_at	PIK3R5	phosphoinositide-3-kinase, regulatory subunit 5	I	8	
221653_x_at	APOL2	apolipoprotein L, 2	I	6	
218608_at	ATP13A2	ATPase type 13A2	D	6	
214149_s_at	ATP6V0E1	ATPase, H+ transporting, lysosomal 9 kDa, V0 subunit e1	I	6	
202017_at	EPHX1	epoxide hydrolase 1, microsomal (xenobiotic)	D	6	yes
239099_at	GCOM1	GRINL1A complex locus 1	I	6	
201185_at	HTRA1	Htra serine peptidase 1	D	6	yes
39402_at	IL1B	interleukin 1, beta	I	6	yes
211354_s_at	LEPR	leptin receptor	D	6	yes
218656_s_at	LHFP	lipoma HMGIC fusion partner	I	6	
236156_at	LIPA	lipase A, lysosomal acid, cholesterol esterase	I	6	
213002_at	MARCKS	myristoylated alanine-rich protein kinase C substrate	I	6	yes
230699_at	PGLS	6-phosphogluconolactonase	I	6	
222176_at	PTEN	phosphatase and tensin homolog	I	6	yes
216153_x_at	RECK	reversion-inducing-cysteine-rich protein with kazal motifs	I	6	yes
200671_s_at	SPTBN1	spectrin, beta, non-erythrocytic 1	D	6	yes
202688_at	TNFSF10	tumor necrosis factor (ligand) superfamily, member 10	I	6	yes
203504_s_at;	ABCA1	ATP-binding cassette, sub-family A (ABC1), member 1	I	4	
203505_at					
241631_at	ARHGEF40	Rho guanine nucleotide exchange factor (GEF) 40	I	4	
220168_at	CASC1	cancer susceptibility candidate 1	I	4	
219799_s_at	DHRS9	dehydrogenase/reductase (SDR family) member 9	I	4	
244642_at	DISC1	disrupted in schizophrenia 1	I	4	
204211_x_at	EIF2AK2	eukaryotic translation initiation factor 2-alpha kinase 2	I	4	yes
231247_s_at	LOC727820	uncharacterized LOC727820	I	4	
242117_at	MAP3K3	mitogen-activated protein kinase kinase kinase 3	I	4	yes
205017_s_at	MBNL2	muscleblind-like splicing regulator 2	D	4	
1553575_at	MT-ND6	mitochondrially encoded NADH dehydrogenase 6	I	4	
217334_at	OR2J3	olfactory receptor, family 2, subfamily J, member 3	D	4	
1565597_at	RBM47	RNA binding motif protein 47	I	4	
227633_at	RHEB	Ras homolog enriched in brain	D	4	yes
228248_at	RICTOR	RPTOR independent companion of MTOR, complex 2	I	4	
243271_at;	SAMD9L	sterile alpha motif domain containing 9-like	I	4	
230036_at					
206995_x_at	SCARF1	scavenger receptor class F, member 1	I	4	
213119_at	SLC36A1	solute carrier family 36 (proton/amino acid symporter), member 1	I	4	
232375_at	STAT1	signal transducer and activator of transcription 1, 91 kDa	I	4	yes

TABLE 8-continued

Complete list of genes differentially expressed in the discovery cohort overlapping between the intra-subject and inter-subject analyses (n = 246).					
Probe set ID	Gene Symbol	Gene Name	Change	Total CFG Score	Evidence or possible roles in apoptosis
236879_at	UBA6	ubiquitin-like modifier activating enzyme 6	I	4	
1563075_s_at	ZC3HAV1	zinc finger CCCH-type, antiviral 1	I	4	
213736_at	COX5B	cytochrome c oxidase subunit Vb	I	3	
203874_s_at	SMARCA1	SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily a, member 1	I	3	
229577_at	AGPAT6	1-acylglycerol-3-phosphate O-acyltransferase 6 (lysophosphatidic acid acyltransferase, zeta)	D	2	
206513_at	AIM2	absent in melanoma 2	I	2	yes
227438_at	ALPK1	alpha-kinase 1	I	2	
210873_x_at	APOBEC3A	apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like 3A	I	2	
239002_at	ASPM	asp (abnormal spindle) homolog, microcephaly associated (<i>Drosophila</i>)	D	2	
222840_at	ATG2B	autophagy related 2B	D	2	
220237_at	ATG3	autophagy related 3	I	2	
211852_s_at	ATRN	attractin	D	2	
243839_s_at	ATXN2	ataxin 2	I	2	yes
204516_at	ATXN7	ataxin 7	I	2	
203140_at; 228758_at	BCL6	B-cell CLL/lymphoma 6	I	2	yes
219072_at	BCL7C	B-cell CLL/lymphoma 7C	D	2	
214068_at	BEAN1	brain expressed, associated with NEDD4, 1	D	2	
212563_at	BOP1	block of proliferation 1	D	2	
233809_at	C15orf63	chromosome 15 open reading frame 63	I	2	yes
221954_at	C20orf111	chromosome 20 open reading frame 111	I	2	yes
1564276_at	C5orf56	chromosome 5 open reading frame 56	I	2	
1553329_at	C7orf45	chromosome 7 open reading frame 45	I	2	
227364_at	CAPZA1	capping protein (actin filament) muscle Z-line, alpha 1	I	2	yes
213596_at	CASP4	caspace 4, apoptosis-related cysteine peptidase	I	2	yes
207500_at	CASP5	caspace 5, apoptosis-related cysteine peptidase	I	2	yes
205099_s_at	CCR1	chemokine (C-C motif) receptor 1	I	2	yes
1554283_at	CCRN4L	CCR4 carbon catabolite repression 4-like (<i>S. cerevisiae</i>)	I	2	
206485_at	CD5	CD5 molecule	D	2	yes
243931_at	CD58	CD58 molecule	I	2	yes
211189_x_at	CD84	CD84 molecule	D	2	
234255_at	CDC42SE2	CDC42 small effector 2	I	2	
209498_at	CEACAM1	carcinoembryonic antigen-related cell adhesion molecule 1 (biliary glycoprotein)	I	2	yes
224198_at	CELA1	chymotrypsin-like elastase family, member 1	D	2	
210069_at	CHKB-CPT1B	CHKB-CPT1B readthrough (non-protein coding)	I	2	
222174_at	CHURC1-FNTB	CHURC1-FNTB readthrough	D	2	
209571_at	CIR1	corepressor interacting with RBPJ, 1	I	2	
219859_at	CLEC4E	C-type lectin domain family 4, member E	I	2	
221698_s_at	CLEC7A	C-type lectin domain family 7, member A	I	2	yes
200861_at	CNOT1	CCR4-NOT transcription complex, subunit 1	D	2	

TABLE 8-continued

Complete list of genes differentially expressed in the discovery cohort overlapping between the intra-subject and inter-subject analyses (n = 246).					
Probe set ID	Gene Symbol	Gene Name	Change	Total CFG Score	Evidence or possible roles in apoptosis
211141_s_at	CNOT3	CCR4-NOT transcription complex, subunit 3	D	2	
1569703_a_at	CORO1C	coronin, actin binding protein, 1C	I	2	yes
205624_at	CPA3	carboxypeptidase A3 (mast cell)	I	2	
203532_x_at	CUL5	cullin 5	D	2	yes
202434_s_at	CYP1B1	cytochrome P450, family 1, subfamily B, polypeptide 1	D	2	yes
208281_x_at	DAZ1	deleted in azoospermia 1	I	2	
209782_s_at	DBP	D site of albumin promoter (albumin D-box) binding protein	D	2	yes
218943_s_at	DDX58	DEAD box polypeptide 58	I	2	yes
240358_at	DENND3	DENN/MADD domain containing 3	I	2	
1556769_a_at	DLGAP1	discs, large (<i>Drosophila</i>) homolog-associated protein 1	I	2	
233052_at	DNAH8	dynein, axonemal, heavy chain 8	D	2	yes
223371_s_at	DNAJC4	DnaJ (Hsp40) homolog, subfamily C, member 4	D	2	
237311_at	DOCK1	dedicator of cytokinesis 1	D	2	yes
244840_x_at	DOCK4	dedicator of cytokinesis 4	I	2	
230207_s_at	DOCK5	dedicator of cytokinesis 5	I	2	
225415_at	DTX3L	deltex 3-like (<i>Drosophila</i>)	I	2	
210525_x_at	EFCAB11	EF-hand calcium binding domain 11	I	2	
214313_s_at	EIF5B	eukaryotic translation initiation factor 5B	I	2	
224727_at	EMC10	ER membrane protein complex subunit 10	D	2	
217245_at	EPAG	early lymphoid activation protein	D	2	
220874_at	EPB41	erythrocyte membrane protein band 4.1 (elliptocytosis 1, RH-linked)	I	2	
210870_s_at	EPM2A	epilepsy, progressive myoclonus type 2A, Lafora disease (laforin)	D	2	yes
239979_at	EPSTI1	epithelial stromal interaction 1 (breast)	I	2	
1570371_a_at	EPT1	ethanolaminephosphotransferase 1 (CDP-ethanolamine-specific)	D	2	
227016_at	ERICH1	glutamate-rich 1	I	2	
225764_at	ETV6	ets variant 6	I	2	yes
214285_at	FABP3	fatty acid binding protein 3, muscle and heart (mammary-derived growth inhibitor)	I	2	
1557385_at	FAM161A	family with sequence similarity 161, member A	D	2	
229543_at	FAM26F	family with sequence similarity 26, member F	I	2	
216950_s_at	FCGR1A	Fc fragment of IgG, high affinity 1a, receptor (CD64)	I	2	yes
1554360_at; 231302_at	FCHSD2	FCH and double SH3 domains 2	I	2	
1553906_s_at	FGD2	FYVE, RhoGEF and PH domain containing 2	I	2	yes
224002_s_at	FKBP7	FK506 binding protein 7	D	2	
211454_x_at; 224288_x_at	FKSG49	FKSG49	I	2	
226419_s_at	FLJ44342	uncharacterized LOC645460	I	2	
228768_at	FNIP1	folliculin interacting protein 1	I	2	
1556667_at	FONG	uncharacterized LOC348751	D	2	
242938_s_at	FOXK2	forkhead box K2	D	2	
230645_at	FRMD3	FERM domain containing 3	I	2	
230744_at	FSTL1	folliculin-like 1	D	2	
1563509_at; 224148_at	FYB	FYN binding protein	I	2	
209416_s_at	FZR1	fizzy/cell division cycle 20 related 1 (<i>Drosophila</i>)	D	2	yes
202748_at; 242907_at	GBP2	guanylate binding protein 2, interferon-inducible	I	2	
229625_at	GBP5	guanylate binding protein 5	I	2	

TABLE 8-continued

Complete list of genes differentially expressed in the discovery cohort overlapping between the intra-subject and inter-subject analyses (n = 246).					
Probe set ID	Gene Symbol	Gene Name	Change	Total CFG Score	Evidence or possible roles in apoptosis
211060_x_at	GPAAL1	glycosylphosphatidylinositol anchor attachment protein 1 homolog (yeast)	D	2	
237690_at	GPR115	G protein-coupled receptor 115	I	2	
218468_s_at	GREM1	gremlin 1	I	2	yes
235957_at	GRIP1	glutamate receptor interacting protein 1	I	2	
213826_s_at	H3F3B	H3 histone, family 3B (H3.3B)	I	2	
205221_at	HGD	homogentisate 1,2-dioxygenase	I	2	
227614_at	HKDC1	hexokinase domain containing 1	D	2	
210747_at	HLA-DQB1	major histocompatibility complex, class II, DQ beta 1	I	2	yes
242001_at	IDH1	isocitrate dehydrogenase 1 (NADP+), soluble	I	2	
226757_at	IFIT2	interferon-induced protein with tetratricopeptide repeats 2	I	2	yes
229450_at	IFIT3	interferon-induced protein with tetratricopeptide repeats 3	I	2	
230128_at	IGLL5	immunoglobulin lambda-like polypeptide 5	I	2	
225025_at	IGSF8	immunoglobulin superfamily, member 8	D	2	
1562468_at	IL1RAP	interleukin 1 receptor accessory protein	I	2	yes
207688_s_at	INHBC	inhibin, beta C	I	2	yes
238725_at	IRF1	interferon regulatory factor 1	I	2	yes
210119_at;	KCNJ15	potassium inwardly-rectifying channel, subfamily J, member 15	I	2	
216782_at	KCNJ2	potassium inwardly-rectifying channel, subfamily J, member 2	I	2	
231513_at;	KCNJ2	potassium inwardly-rectifying channel, subfamily J, member 2	I	2	
206765_at	KIAA0494	KIAA0494	I	2	
1559023_a_at	KIAA1967	KIAA1967	D	2	yes
225193_at	KLHDC3	kelch domain containing 3	D	2	
208784_s_at	KPNA3	karyopherin alpha 3 (importin alpha 4)	I	2	
208974_x_at	KPNB1	karyopherin (importin) beta 1	I	2	yes
1555384_a_at	LARP4	La ribonucleoprotein domain family, member 4	D	2	
215229_at	LOC100129973	uncharacterized LOC100129973	D	2	
1569746_s_at	LOC100505783	uncharacterized LOC100505783	I	2	
215322_at	LONRF1	LON peptidase N-terminal domain and ring finger 1	I	2	
233818_at	LTN1	listerin E3 ubiquitin protein ligase 1	I	2	
232283_at	LYSMD1	LysM, putative peptidoglycan-binding, domain containing 1	I	2	
215902_at	MARCH 6	membrane-associated ring finger (C3HC4) 6, E3 ubiquitin protein ligase	I	2	
1554730_at	MCTP1	multiple C2 domains, transmembrane 1	I	2	
235589_s_at	MDM4	Mdm4 p53 binding protein homolog (mouse)	I	2	yes
222567_s_at	MEX3C	mex-3 homolog C (<i>C. elegans</i>)	D	2	
241541_at	MIB2	mindbomb E3 ubiquitin protein ligase 2	I	2	
225826_at	MMAB	methylmalonic aciduria (cobalamin deficiency) cblB type	D	2	
239273_s_at	MMP28	matrix metalloproteinase 28	D	2	yes
221995_s_at	MRP63	mitochondrial ribosomal protein 63	I	2	
228846_at	MXD1	MAX dimerization protein 1	I	2	yes
211010_s_at	NCR3	natural cytotoxicity triggering receptor 3	D	2	yes
243357_at	NEGR1	neuronal growth regulator 1	D	2	
223218_s_at	NFKBIZ	nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, zeta	I	2	yes
214101_s_at	NPEPPS	aminopeptidase puromycin sensitive	I	2	

TABLE 8-continued

Complete list of genes differentially expressed in the discovery cohort overlapping between the intra-subject and inter-subject analyses (n = 246).					
Probe set ID	Gene Symbol	Gene Name	Change	Total CFG Score	Evidence or possible roles in apoptosis
1557071_s_at	NUB1	negative regulator of ubiquitin-like proteins 1	I	2	yes
1561847_at	NUDT17	nudix (nucleoside diphosphate linked moiety X)-type motif 17	D	2	
1569990_at	NUDT3	nudix (nucleoside diphosphate linked moiety X)-type motif 3	I	2	
243934_at	ODF3B	outer dense fiber of sperm tails 3B	I	2	
229787_s_at	OGT	O-linked N-acetylglucosamine (GlcNAc) transferase	I	2	yes
1569617_at	OSBP2	oxysterol binding protein 2	D	2	
243287_s_at	OSTM1	osteopetrosis associated transmembrane protein 1	I	2	
231838_at	PABPC1L	poly(A) binding protein, cytoplasmic 1-like	I	2	
235157_at	PARP14	poly (ADP-ribose) polymerase family, member 14	I	2	
227807_at	PARP9	poly (ADP-ribose) polymerase family, member 9	I	2	
241956_at	PCGF5	polycomb group ring finger 5	I	2	
222045_s_at	PCIF1	PDX1 C-terminal inhibiting factor 1	D	2	
217695_x_at	PELI1	pellino E3 ubiquitin protein ligase 1	I	2	
225958_at	PHC1	polyhomeotic homolog 1 (<i>Drosophila</i>)	I	2	
237867_s_at	PID1	phosphotyrosine interaction domain containing 1	D	2	
216112_at	PKN2	protein kinase N2	I	2	yes
241916_at	PLSCR1	phospholipid scramblase 1	I	2	yes
235508_at	PML	promyelocytic leukemia	I	2	yes
202884_s_at	PPP2R1B	protein phosphatase 2, regulatory subunit A, beta	D	2	yes
1559119_at	PPP6R3	protein phosphatase 6, regulatory subunit 3	I	2	
221270_s_at	QTRT1	queuine tRNA-ribosyltransferase 1	D	2	
241320_at	R3HDM1	R3H domain containing 1	I	2	
1553285_s_at	RAD9B	RAD9 homolog B (<i>S. pombe</i>)	I	2	
202052_s_at	RAI14	retinoic acid induced 14	D	2	yes
230466_s_at	RASSF3	Ras association (RalGDS/AF-6) domain family member 3	I	2	
204927_at	RASSF7	Ras association (RalGDS/AF-6) domain family (N-terminal) member 7	D	2	
237626_at	RB1CC1	RB1-inducible coiled-coil 1	I	2	yes
232150_at	RBCK1	RanBP-type and C3HC4-type zinc finger containing 1	I	2	yes
1560340_s_at	RP9P	retinitis pigmentosa 9 pseudogene	I	2	
214041_x_at	RPL37A	ribosomal protein L37a	I	2	
200908_s_at	RPLP2	ribosomal protein, large, P2	I	2	
242625_at	RSAD2	radical S-adenosyl methionine domain containing 2	I	2	
214370_at	S100A8	S100 calcium binding protein A8	I	2	yes
242190_at	SDAD1	SDA1 domain containing 1	I	2	
214257_s_at	SEC22B	SEC22 vesicle trafficking protein homolog B (<i>S. cerevisiae</i>) (gene/pseudogene)	I	2	
223121_s_at	SFRP2	secreted frizzled-related protein 2	D	2	yes
35626_at	SGSH	N-sulfoglucosamine sulfohydrolase	D	2	
228527_s_at	SLC25A37	solute carrier family 25 (mitochondrial iron transporter), member 37	I	2	
234268_at	SLC2A13	solute carrier family 2 (facilitated glucose transporter), member 13	I	2	
235536_at	SNORD89	small nucleolar RNA, C/D box 89	I	2	
208012_x_at; 209762_x_at	SP110	SP110 nuclear body protein	I	2	

TABLE 8-continued

Complete list of genes differentially expressed in the discovery cohort overlapping between the intra-subject and inter-subject analyses (n = 246).					
Probe set ID	Gene Symbol	Gene Name	Change	Total CFG Score	Evidence or possible roles in apoptosis
228975_at	SP6	Sp6 transcription factor	D	2	
1557593_at	SPAG17	sperm associated antigen 17	D	2	
202523_s_at	SPOCK2	sparc/osteonectin, ewcv and kazal-like domains proteoglycan (testican) 2	D	2	
243522_at	SPPL3	signal peptide peptidase like 3	I	2	
213562_s_at	SQLE	squalene epoxidase	D	2	
219055_at	SRBD1	S1 RNA binding domain 1	I	2	
1565566_a_at	STX7	syntaxin 7	I	2	
1557305_at	TACC1	transforming, acidic coiled-coil containing protein 1	I	2	
216226_at	TAF4B	TAF4b RNA polymerase II, TATA box binding protein (TBP)-associated factor, 105 kDa	D	2	
231193_s_at	TAOK1	TAO kinase 1	I	2	yes
225973_at	TAP2	transporter 2, ATP-binding cassette, sub-family B (MDR/TAP)	I	2	
221398_at	TAS2R8	taste receptor, type 2, member 8	I	2	
213401_s_at	TBL1X	transducin (beta)-like 1X-linked	I	2	
1566208_at	TCEA1	transcription elongation factor A (SII), 1	I	2	
1552804_a_at	TIRAP	toll-interleukin 1 receptor (TIR) domain containing adaptor protein	D	2	yes
224321_at	TMEFF2	transmembrane protein with EGF-like and two follistatin-like domains 2	I	2	
235159_at	TMEM140; 243465_at	transmembrane protein 140	I	2	
238063_at	TMEM154	transmembrane protein 154	I	2	
227386_s_at	TMEM200B	transmembrane protein 200B	I	2	
1554206_at	TMLHE	trimethyllysine hydroxylase, epsilon	I	2	
206025_s_at; 206026_s_at	TNFAIP6	tumor necrosis factor, alpha-induced protein 6	I	2	
1555557_a_at	TNK2	tyrosine kinase, non-receptor, 2	D	2	
1558354_s_at	TOP1	topoisomerase (DNA) I	I	2	yes
231978_at	TPCN2	two pore segment channel 2	I	2	
223599_at	TRIM6	tripartite motif containing 6	I	2	
242688_at	TRIP12	thyroid hormone receptor interactor 12	I	2	
1565887_at	TRPM7	transient receptor potential cation channel, subfamily M, member 7	I	2	yes
215107_s_at	TTC22	tetratricopeptide repeat domain 22	D	2	
202476_s_at	TUBGCP2	tubulin, gamma complex associated protein 2	D	2	yes
228588_s_at	UBE2B	ubiquitin-conjugating enzyme E2B	I	2	yes
1568903_at	UBR5	ubiquitin protein ligase E3 component n-recogin 5	I	2	yes
205586_x_at	VGf	VGf nerve growth factor inducible	D	2	
242390_at	WDFY1	WD repeat and FYVE domain containing 1	I	2	
201421_s_at	WDR77	WD repeat domain 77	D	2	
1569428_at	WIBG	within bgcn homolog (<i>Drosophila</i>)	D	2	yes
213734_at	WSB2	WD repeat and SOCS box containing 2	I	2	
228617_at	XAF1	XIAP associated factor 1	I	2	yes
1554037_a_at	ZBTB24	zinc finger and BTB domain containing 24	D	2	
219062_s_at	ZCCHC2	zinc finger, CCHC domain containing 2	I	2	
1555982_at	ZFYVE16	zinc finger, FYVE domain containing 16	I	2	
228864_at	ZNF653	zinc finger protein 653	D	2	

[0098] CD24, the top biomarker decreased in suicidal subjects of this Example, also has roles in apoptosis. Mice lacking CD24 show an increased rate of apoptosis (Duckworth C. A. et al., "CD24 is expressed in gastric parietal cells and regulates apoptosis and the response to *Helicobacter felis* infection in the murine stomach," American Journal of Physiology, Gastrointestinal and Liver Physiology 303, G915-926, doi:10.1152/ajpgi.00068.2012 (2012)). It could be that simpler mechanisms related to cellular survival and programmed cell-death decision have been recruited by evolution for higher mental functions such as thoughts and behaviors leading to suicidality. In that sense, suicidality could be viewed as whole-organism self-poptosis. Interestingly, lithium, a medication with clinical evidence for preventing suicidality in bipolar disorder, has anti-apoptotic effects at a cellular level. Imaging studies have shown reduced gray matter volume in the brain of individuals with bipolar disorder and history of suicide attempts. Long-term lithium treatment was associated with increased gray matter volumes in the same areas where suicide was associated with decreased gray matter.

[0099] Taken together, the results of this Example have implications for the understanding of suicide, as well as for the development of objective laboratory tests and tools to diagnose and track suicidal risk and to monitor response to treatment.

[0100] More particularly, it was found that suicidality may be associated with dysphoric mood, as well as increased psychosis, anxiety and stress. SAT1 blood gene expression levels, in particular, showed a trend towards increase in low mood, high psychosis, high anxiety, and high stress in the bipolar subjects (see FIGS. 4A-4F).

Example 2

[0101] In this Example, SAT1 was validated by analyzing subsequent hospitalizations with and without suicidality and to previous hospitalizations with and without suicidality in two live follow-up cohorts, one bipolar (n=42) and one psychosis (schizophrenia/schizoaffective; n=46).

[0102] Particularly, the bipolar follow-up cohort (Table 9A) consisted of male Caucasian subjects in which whole-genome blood gene expression data, including levels of SAT1, were obtained at the testing visits as described in Example 1. If the subjects had multiple testing visits, the visit with the highest SAT1 level was selected for this analysis. The subjects' subsequent number of hospitalizations with or without suicidality was tabulated from electronic medical records.

[0103] The psychosis (schizophrenia/schizoaffective) follow-up cohort (n=46) (Table 9B) similarly consisted of Caucasian subjects in which whole-genome blood gene expression data, including levels of SAT1, were obtained at testing visits as described for the bipolar follow-up cohort. If the subjects had multiple testing visits, the visit with the highest SAT1 level was selected for this analysis. The subjects' subsequent number of hospitalizations with or without suicidality was tabulated from electronic medical records. A hospitalization was deemed to be without suicidality if suicidality was not listed as a reason for admission, and no suicidal ideation was described in the admission and discharge medical notes. Conversely, a hospitalization was deemed to be due to suicidality if suicidal acts or intent was listed as a reason for admission, and suicidal ideation was described in the admission and discharge medical notes.

TABLE 9A

Demographic Data for Live Bipolar Cohort (n = 42)										
SubjectID-Visit	Diagnosis	Age	Gender	Ethnicity	SAT1 Levels	Years since testing	Future Hosp. w/o suicidality	Future Hosp. due to suicidality	Frequency of Future Hosp. w/o suicidality	Frequency of Future Hosp. due to suicidality
phchp234v1	Bipolar II Disorder	44	M	Caucasian	1955.2	0.83	0	0	0	0
phchp053v2	Bipolar I Disorder	58	M	Caucasian	2178.3	5.67	4	0	0.71	0
phchp152v1	Bipolar I Disorder	45	M	Caucasian	2178.8	2.33	0	0	0	0
phchp122v1	Bipolar Disorder NOS	51	M	Caucasian	2245.6	0.58	0	0	0	0
phchp190v3	Bipolar Disorder NOS	50	M	Caucasian	2300.6	1.25	0	0	0	0
phchp020v3	Bipolar Disorder NOS	63	M	Caucasian	2342.6	4.08	0	0	0	0
phchp113v1	Bipolar I Disorder	37	M	Caucasian	2437.4	3.00	0	0	0	0
phchp132v2	Bipolar I Disorder	51	M	Caucasian	2558.9	2.33	0	0	0	0
phchp184v3	Bipolar Disorder NOS	64	M	Caucasian	2575.4	1.33	0	0	0	0
phchp039v3	Bipolar I Disorder	52	M	Caucasian	2580.1	5.75	0	0	0	0
phchp147v1	Bipolar II Disorder	38	M	Caucasian	2582.8	2.25	0	0	0	0
phchp178v1	Bipolar I Disorder	49	M	Caucasian	2616.8	1.0	0	0	0	0
phchp136v3	Bipolar I Disorder	41	M	Caucasian	2635.9	2.0	0	0	0	0
phchp045v1	Bipolar I Disorder	36	M	Caucasian	2721.0	5.42	0	0	0	0
phchp224v1	Bipolar I Disorder	59	M	Caucasian	2748.1	1.08	1	1	0.92	0.92

TABLE 9A-continued

Demographic Data for Live Bipolar Cohort (n = 42)										
SubjectID- Visit	Diagnosis	Age	Gender	Ethnicity	SAT1 Levels	Years since testing	Future Hosp. w/o suicidality	Future Hosp. due to suicidality	Frequency of Future Hosp. w/o suicidality	Frequency of Future Hosp. due to suicidality
phchp183v1	Bipolar I Disorder	48	M	Caucasian	2750.9	0.42	2	1	4.80	2.40
phchp171v2	Bipolar Disorder NOS	36	M	Caucasian	2795.7	1.50	0	0	0	0
phchp166v1	Bipolar Disorder NOS	56	M	Caucasian	2829.6	1.92	0	0	0	0
phchp253v1	Bipolar Disorder NOS	25	M	Caucasian	2888.5	1.0	0	0	0	0
phchp186v1	Bipolar II Disorder	43	M	Caucasian	2901.5	1.67	0	0	0	0
phchp079v2	Bipolar Disorder	44	M	Caucasian	3053.2	4.50	0	0	0	0
phchp128v1	Bipolar I Disorder	45	M	Caucasian	3118.6	2.67	0	0	0	0
phchp080v1	Bipolar I Disorder	44	M	Caucasian	3153.6	5.00	0	0	0	0
phchp088v1	Bipolar I Disorder	44	M	Caucasian	3194.1	4.58	0	10	0	2.18
phchp109v1	Bipolar I Disorder	22	M	Caucasian	3200.8	3.00	1	2	0.33	0.67
phchp134v3	Bipolar II Disorder	59	M	Caucasian	3202.3	1.92	0	0	0	0
phchp153v1	Bipolar II Disorder	55	M	Caucasian	3304.9	2.0	0	0	0	0
phchp274v2	Bipolar Disorder NOS	48	M	Caucasian	3349.0	0.50	0	0	0	0
phchp140v3	Bipolar II Disorder	38	M	Caucasian	3393.8	1.92	0	0	0	0
phchp030v3	Bipolar I Disorder	49	M	Caucasian	3395.2	5.92	0	3	0	0.51
phchp124v1	Bipolar I Disorder	53	M	Caucasian	3660.9	2.50	0	6	0	2.40
phchp095v3	Bipolar I Disorder	29	M	Caucasian	3695.4	0.33	0	1	0	3.00
phchp100v1	Bipolar I Disorder	28	M	Caucasian	3767.8	1.58	0	0	0	0
phchp210v3	Bipolar I Disorder	44	M	Caucasian	3844.6	0.50	0	0	0	0
phchp219v1	Bipolar Disorder NOS	61	M	Caucasian	3845.1	1.17	0	0	0	0
phchp031v3	Bipolar I Disorder	52	M	Caucasian	4080.7	4.08	1	0	0.24	0
phchp093v3	Bipolar I Disorder	52	M	Caucasian	4137.4	2.67	0	1	0	0.38
phchp067v1	Bipolar II Disorder	39	M	Caucasian	4214.7	5.58	0	0	0	0
phchp142v3	Bipolar I Disorder	55	M	Caucasian	4310.7	1.92	0	0	0	0
phchp112v2	Bipolar I Disorder	46	M	Caucasian	4410.4	1.33	0	0	0	0
phchp149v2	Bipolar Disorder NOS	45	M	Caucasian	4586.9	2.00	1	0	0.5	0
phchp117v1	Bipolar I Disorder	43	M	Caucasian	6531.1	3.00	0	0	0	0

TABLE 9B

Demographic Data for Live Psychosis Cohort (n = 46)										
SubjectID- Visit	Diagnosis	Age	Gender	Ethnicity	SAT1 Levels	Years since testing	Future Hosp. w/o suicidality	Future Hosp. due to suicidality	Frequency of Future Hosp. w/o suicidality	Frequency of Future Hosp. due to suicidality
phchp222v2	Schizophrenia	60	M	Caucasian	1410.6	0.67	0	0	0	0
phchp175v1	Schizoaffective Disorder	42	M	Caucasian	1773.9	2.08	0	0	0	0
phchp139v1	Schizophrenia	24	M	Caucasian	1774.6	0.25	0	0	0	0

TABLE 9B-continued

Demographic Data for Live Psychosis Cohort (n = 46)										
SubjectID- Visit	Diagnosis	Age	Gender	Ethnicity	SAT1 Levels	Years since testing	Future Hosp. w/o suicidality	Future Hosp. due to suicidality	Frequency of Future Hosp. w/o suicidality	Frequency of Future Hosp. due to suicidality
phchp025v1	Schizophrenia	42	M	Caucasian	2004.6	6.83	0	0	0	0
phchp051v1	Schizoaffective Disorder	52	M	Caucasian	2083.8	5.83	0	0	0	0
phchp148v1	Schizophrenia	25	M	Caucasian	2254.7	2.17	1	0	0.46	0
phchp133v1	Schizophrenia	55	M	Caucasian	2286	2.75	0	2	0	0.73
phchp033v1	Schizoaffective Disorder	48	M	Caucasian	2291.4	2.58	0	1	0	0.39
phchp027v1	Schizoaffective Disorder	40	M	Caucasian	2406.3	6.67	3	0	0.45	0
phchp012v1	Schizoaffective Disorder	55	M	Caucasian	2458.1	5.17	1	1	0.19	0.19
phchp089v2	Schizoaffective Disorder	33	M	Caucasian	2545.3	4.42	0	0	0	0
phchp060v1	Schizophrenia	62	M	Caucasian	2589.2	3.50	2	0	0.57	0
phchp046v1	Schizoaffective Disorder	45	M	Caucasian	2732.3	6.17	0	1	0	0.16
phchp103v1	Schizoaffective Disorder	61	M	Caucasian	2763.7	2.58	1	2	0.39	0.77
phchp010v2	Schizoaffective Disorder	45	M	Caucasian	2778.5	6.92	0	0	0	0
phchp005v1	Schizoaffective Disorder	45	M	Caucasian	2797.8	7.33	1	1	0.14	0.14
phchp022v1	Schizophrenia	48	M	Caucasian	2846.6	6.83	0	0	0	0
phchp195v3	Schizophrenia	53	M	Caucasian	2846.6	1.17	0	0	0	0
phchp129v1	Schizoaffective Disorder	22	M	Caucasian	2871.5	2.83	5	1	1.76	0.35
phchp120v1	Delusional Disorder	51	M	Caucasian	2877.9	3.00	0	0	0	0
phchp211v1	Schizophrenia	62	M	Caucasian	2879.9	1.25	0	0	0	0
phchp277v2	Schizophrenia	50	M	Caucasian	2904.8	0.58	0	0	0	0
phchp101v1	Schizoaffective Disorder	74	M	Caucasian	2923.7	3.67	0	1	0	0.27
phchp116v1	Schizoaffective Disorder	47	M	Caucasian	2962.1	0.50	0	1	0	2.00
phchp052v1	Schizophrenia	60	M	Caucasian	2989.9	0.83	0	0	0	0
phchp090v3	Schizophrenia	24	M	Caucasian	3046.4	1.00	0	2	0	2.00
phchp197v1	Schizophrenia	56	M	Caucasian	3046.6	1.67	1	0	0.60	0
phchp061v3	Schizophrenia	50	M	Caucasian	3115.6	4.92	1	6	0.20	1.22
phchp057v1	Schizoaffective Disorder	47	M	Caucasian	3233.8	5.92	0	0	0	0
phchp105v2	Schizoaffective Disorder per chip	59	M	Caucasian	3297.6	2.83	2	0	0.71	0
phchp087v3	Schizoaffective Disorder	66	M	Caucasian	3523.5	4.25	0	0	0	0
phchp091v1	Schizoaffective Disorder	55	M	Caucasian	3534.5	4.75	0	0	0	0
phchp069v3	Schizophrenia	48	M	Caucasian	3819.8	5.25	0	0	0	0
phchp062v3	Schizophrenia	57	M	Caucasian	3878.8	5.42	0	0	0	0
phchp099v2	Schizophrenia	49	M	Caucasian	3993.4	3.58	0	0	0	0
phchp049v1	Schizoaffective Disorder	46	M	Caucasian	4012.3	6.08	0	0	0	0
phchp040v3	Schizoaffective Disorder	50	M	Caucasian	4019.2	5.25	1	0	0.19	0
phchp042v3	Schizoaffective Disorder	44	M	Caucasian	4124.5	5.50	0	0	0	0
phchp075v3	Schizoaffective Disorder	58	M	Caucasian	4127.1	4.83	1	5	0.21	1.03
phchp108v2	Schizophrenia	42	M	Caucasian	4231.9	3.17	0	0	0	0
phchp085v3	Schizoaffective Disorder	57	M	Caucasian	4335.9	4.50	0	0	0	0
phchp151v3	Schizophrenia	24	M	Caucasian	4390.9	2.00	1	1	0.50	0.50
phchp065v3	Schizoaffective Disorder	62	M	Caucasian	4439.2	5.25	0	0	0	0
phchp086v3	Schizophrenia	49	M	Caucasian	4545.4	4.25	0	0	0	0
phchp073v3	Schizoaffective Disorder	65	M	Caucasian	4874.4	4.92	0	12	0	2.44
phchp072v3	Schizoaffective Disorder	60	M	Caucasian	5911.1	5.08	0	1	0	0.20

[0104] For future hospitalization analyses, robust multi-array analysis (RMA) as described in Example 1 was conducted and normalized for each cohort, prior to looking at biomarker levels in individual subjects. One-tail t-tests with equal variance were used for statistical comparisons. ROC curves were calculated using SPSS software for each of the four-dimensional analyses, predicting the state variable of hospitalizations due to suicidality.

[0105] Higher SAT1 levels compared to lower SAT1 levels at time of testing differentiated future as well as past hospitalizations due to suicidality in the bipolar disorder subjects (FIGS. 5A-5E). A similar, but weaker, pattern was exhibited in the psychosis (schizophrenia/schizoaffective) subjects (FIGS. 6A-6E). Remarkably, besides SAT1, three other biomarkers (PTEN, MARCKS and MAP3K3) of the six biomarkers that survived Bonferroni correction in the suicide completers cohort validation step also showed similar but weaker results (Table 10 and FIGS. 7A-7C).

TABLE 10

Prospective and Retrospective Differentiation of Hospitalizations and Suicidality								
	Bipolar Disorder (n = 42)				Psychosis (n = 46) Schizophrenia/Schizoaffective			
	Future Hospitalizations (since testing)		Past Hospitalizations (prior to testing)		Future Hospitalizations (since testing)		Past Hospitalizations (prior to testing)	
	Without	With Suicidality	Without	With Suicidality	Without	With Suicidality	Without	With Suicidality
SAT1	NS	H: 0.1195 T: 0.0484	NS	H: 0.0743 T: 0.0363	NS	NS (H: 0.0519)	NS	H: 0.0274 T: 0.0742
PTEN	NS	H: 0.0271 T: 0.0324	NS	H: 0.0598 T: 0.0491	NS	NS	NS	NS
MARCKS	NS	NS	NS	H: 0.0227 T: 0.0242	NS	NS	NS	NS
MAP3K3	NS	NS	NS	H: 0.2052 T: 0.0273	NS	NS	NS	NS
UBA6	NS	NS	NS	NS	NS	NS	NS	NS
MT-ND6	NS	NS	NS	NS	NS	NS	NS	NS
Panel of 3 (SAT1, PTEN, MAP3K3)	NS	H: 0.0184 T: 0.0530	NS	H: 0.04905 T: 0.04914	NS	NS	NS	NS
Panel of 6 (SAT1, PTEN, MAP3K3, UBA6, MARCK, MT-ND6)	NS	H: 0.1501 T: 0.0159	NS	H: 0.0728 T: 0.0101	NS	NS	NS	NS

[0106] Taken together, the prospective and retrospective hospitalization data suggests SAT1, PTEN, MARCKS and MAP3K3 may be not only a state marker but perhaps a trait marker as well.

[0107] A multi-dimensional approach was also conducted to predict future hospitalizations, by adding data about mood, anxiety, and psychosis to the data about SAT1 expression levels (FIGS. 8A-8C). The ROC curve improved in a step-wise fashion, from an AUC of 0.640 with SAT1 alone, to an AUC of 0.798 with SAT1 and anxiety, an AUC of 0.813 with SAT1, anxiety and mood, and an AUC of 0.835 with SAT1, anxiety, mood and psychosis. Levels of SAT1 were identified that provided different levels of sensitivity and specificity (Table 11). The anxiety and mood information was obtained from simple visual analog scales, previously described in Niculescu, et al., "PhenoChipping of psychotic disorders: a novel approach for deconstructing and quantitating psychiatric phenotypes. *American Journal of Medical Genetics. Part*

B, Neuropsychiatric genetics: the official publication of the International Society of Psychiatric Genetics 141B, 653-662, doi:10.1002/ajmg.b.30404 (2006).

TABLE 11

SAT1 Expression Level Cut-offs from the ROC Curve (FIGS. 8A-8C)				
Cut-off	SAT1 Expression Levels	Sensitivity	Specificity	Accuracy
Higher Sensitivity	2723.512	100.00%	41.18%	70.59%
Intermediate	3173.874	75.00%	61.76%	68.38%
Higher Specificity	3394.539	50.00%	73.53%	61.77%

[0108] The multi-dimensional approach described above for SAT1 was also conducted to predict future hospitaliza-

tions, by adding data about mood, anxiety, and psychosis to the data about the six top biomarkers' expression levels (BioM 6, including the biomarkers SAT1, PTEN, MARCKS, MAP3K3, UBA6, and MT-ND6; FIGS. 9A-9B).

[0109] These results demonstrate that combining clinical scale data for anxiety and mood with the blood biomarker data improves predictability of increased suicide ideation and/or future hospitalization.

[0110] The psychosis information was based on combining of the scores on the hallucinations and delusions in the PANSS (FIG. 10). Of note, this simple clinical-genomic approach did not directly ask about suicidal ideation, which some individuals may deny or choose not to share with clinicians.

[0111] Using discovery in live subjects and validation in suicide completers, possible biomarkers for suicidality were found. The top biomarker finding, SAT1, as well as PTEN, MARCKS and MAP3K3, were additionally validated by pro-

spective and retrospective analyses in live subjects, looking at ability to predict and differentiate future and past hospitalizations due to suicidality in bipolar disorder and psychosis (schizophrenia/schizoaffective) (Table 10).

[0112] Beyond predictions, as a window into the biology of suicidality, the current Examples show overlap at a gene and pathway level with apoptosis. SAT1, for example, is a key catabolic enzyme for polyamines. Polyamine levels within cells control cell viability, and significant decreases in polyamine levels can result in apoptosis. They seem to reflect an endowment for cellular and organismal activity and growth, key characteristics of mood. SAT1, which is increased in live suicidal ideation subjects and in suicide completers in the Examples, is highly inducible by a variety of stimuli, including toxins, cytokines, heat shock, ischemia, and other stresses. SAT1 overexpressing mice had alterations in their polyamine pool, hair loss, infertility and weight loss (Pietila et al., Activation of polyamine catabolism profoundly alters tissue polyamine pools and affects hair growth and female fertility in transgenic mice overexpressing spermidine/spermine N1-acetyltransferase. *J Biol. Chem.* 272, 18746-18751 (1997); Min et al., Altered levels of growth-related and novel gene transcripts in reproductive and other tissues of female mice overexpressing spermidine/spermine N1-actyltransferase (SSAT). *J. Biol. Chem.* 277, 3647-3657, doi:10.1074/jbc.M100751200 (2002)). Turecki and colleagues have provided compelling evidence for changes in the polyamine system in the brain of suicide completers (Fiori et al., Global gene expression profiling of the polyamine system in suicide completers. *Int. J. Neuropsychopharmacol.* 14, 595-605, doi:10.1017/S1461145710001574 (2011)).

[0113] CD24, the top biomarker found to decrease in suicidal subjects, also has roles in apoptosis. Specifically, mice lacking CD24 showed an increased rate of apoptosis (Duckworth et al. CD24 is expressed in gastric parietal cells and regulates apoptosis and the response to *Helicobacter felis* infection in the murine stomach. *American Journal of Physiology. Gastrointestinal and liver physiology* 303, G915-926, doi:10.1152/ajpgi.00068.2012 (2012)).

[0114] It could be that simpler mechanisms related to cellular survival and programmed cell-death decision have been recruited by evolution for higher mental functions such as feelings, thoughts, actions and behaviors leading to suicidality. In that sense, suicidality could be viewed as whole-organism self-apoptosis. Apoptosis mechanisms have previously been implicated in mood disorders, and their inhibition in affective resilience (Malkesman et al. Targeting the BH3-interacting domain death agonist to develop mechanistically unique antidepressants. *Mol. Psychiatry* 17, 770-780, doi:10.1038/mp.2011.77 (2012)). Interestingly, lithium, a medication with clinical evidence for preventing suicidality in bipolar disorder, has anti-apoptotic effects at a cellular level (Lowthert et al., Increased ratio of anti-apoptotic to pro-apoptotic Bcl2 gene-family members in lithium-responders one month after treatment initiation. *Biology of Mood & Anxiety Disorders* 2, 15, doi:10.1186/2045-5380-2-15 (2012)). Imaging studies have shown reduced gray matter volume in the brain of individuals with bipolar disorder and history of suicide attempts. Long-term lithium treatment was associated with increased gray matter volumes in the same areas where suicide was associated with decreased gray matter (Benedetti et al., Opposite effects of suicidality and lithium on gray matter volumes in bipolar depression. *J Affect Disord* 135, 139-147, doi:10.1016/j.jad.2011.07.006 (2011)).

[0115] In view of the above, it will be seen that the several advantages of the disclosure are achieved and other advantageous results attained. As various changes could be made in the above methods without departing from the scope of the disclosure, it is intended that all matter contained in the above description and shown in the accompanying drawings shall be interpreted as illustrative and not in a limiting sense.

[0116] When introducing elements of the present disclosure or the various versions, embodiment(s) or aspects thereof, the articles "a", "an", "the" and "said" are intended to mean that there are one or more of the elements. The terms "comprising", "including" and "having" are intended to be inclusive and mean that there may be additional elements other than the listed elements.

1-8. (canceled)

9. A method for monitoring response of a subject to a treatment for suicidal risk, the method comprising:

obtaining an expression level of a biomarker from the subject;

administering a treatment for suicidal risk to the subject; and

determining an expression level of the biomarker in a sample obtained from the subject after the treatment is administered, wherein a change in the expression level of the biomarker in the sample obtained from the subject after the treatment is administered as compared to the expression level of the biomarker before the treatment is administered indicates a response to the treatment.

10. The method of claim 9, wherein the response is an increase in expression level of the biomarker.

11. The method of claim 10, wherein the biomarker is selected from the group consisting of small cell lung carcinoma cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof.

12. The method of claim 9, wherein the response is a decrease in expression level of the biomarker.

13. The method of claim 12, wherein the biomarker is selected from the group consisting of spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H⁺ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1

(DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAM9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2), and combinations thereof.

14. The method of claim 9, wherein the treatment for suicidal risk is selected from the group consisting of a drug, a nutritional, and combinations thereof.

15. The method of claim 14, wherein the drug is selected from the group consisting of clozapine, lithium, IL-1 trap, canakinumab, nicorandil, amiodarone, arsenic trioxide, vemurafenib, elsamitucin, T 0128, CT-2106, BN80927, taf-luposide, TAS-103, beta-lapachone, irinotecan, topo tecan, 9-amino-20-camptothecin, rubitecan, gimatecan, karenitecin, and combinations thereof.

16. The method of claim 14, wherein the nutritional is an omega-3 fatty acid.

17. The method of claim 9, wherein the sample obtained from the subject is selected from the group consisting of whole blood, leukocytes, megakaryocytes, brain, cerebrospinal fluid, olfactory epithelium cells, fibroblasts from skin biopsies, induced pluripotent stem cells, and neuronal-like cells derived therefrom.

18. The method of claim 9, wherein the biomarker is selected from the group consisting of spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H⁺ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAM9L); scavenger receptor class F member 1 (SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase

subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2), and combinations thereof in the blood sample of the subject is increased as compared to the reference expression level, and wherein the expression of the blood biomarker selected from the group consisting of cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof in the blood sample of the subject is decreased as compared to the reference expression level.

19. A method for determining suicidal risk as a side-effect of an antidepressant, the method comprising:

obtaining an expression level of a biomarker from a subject;

administering an antidepressant to the subject; and

determining an expression level of the biomarker in a sample obtained from the subject after the antidepressant is administered, wherein a change in the expression level of the biomarker in the sample obtained from the subject after the antidepressant is administered as compared to the expression level of the biomarker before the antidepressant is administered indicates suicidal risk as a side-effect of the antidepressant.

20. The method of claim 19, wherein the antidepressant is selected from the group consisting of bupropion, citalopram, escitalopram, fluoxetine, fluvoxamine, mirtazapine, nefazodone, paroxetine, sertraline, and venlafaxine.

21. The method of claim 19, wherein the biomarker is selected from the group consisting of spermidine/spermine N1-acetyltransferase 1 (SAT1); forkhead box N3 (FOXN3); guanylate binding protein 1 (GBP1); phosphoinositide-3-kinase regulatory subunit 5 (PIK3R5); apolipoprotein L2 (APOL2); ATPase H⁺ transporting lysosomal 9 kDa, V0 subunit e1 (ATP6V0E1); GRINL1A complex locus (GCOM1); interleukin 1 beta (IL1B); lipoma HMGIC fusion partner (LHFP); lipase A (LIPA); myristoylated alanine-rich protein kinase C substrate (MARCKS); 6-phosphogluconolactonase (PGLS); phosphatase and tensin homolog (PTEN); reversion-inducing-cysteine-rich protein with kazal motifs (RECK); tumor necrosis factor (ligand) superfamily member 10 (TNFSF10); ATP-binding cassette, subfamily A (ABC1) member 1 (ABCA1); Rho guanine nucleotide exchange factor (GEF) 40 (ARHGEF4; FLJ10357); cancer susceptibility candidate 1 (CASC1); dehydrogenase/reductase (SDR family) member 9 (DHRS9); disrupted in schizophrenia 1 (DISC1); eukaryotic translation initiation factor 2-alpha kinase 2 (EIF2AK2); uncharacterized LOC727820 (LOC727820); mitogen-activated protein kinase kinase 3 (MAP3K3); mitochondrially encoded NADH dehydrogenase 6 (MT-ND6; ND6); RNA binding motif protein 47 (RBM47); RPTOR independent companion of MTOR complex 2 (RICTOR); sterile alpha motif domain containing 9-like (SAM9L); scavenger receptor class F member 1

(SCARF1); solute carrier family 36 (proton/amino acid symporter) member 1 (SLC36A1); signal transducer and activator of transcription 1, 91 kDa (STAT1); cytochrome c oxidase subunit Vb (COX5B); SWI/SNF related matrix associated actin dependent regulator of chromatin subfamily a member 1 (SMARCA1); ubiquitin-like modifier activating enzyme 6 (UBA6); zinc finger CCCH-type antiviral 1 (ZC3HAV1); tyrosine kinase, non-receptor 2 (TNK2); cluster 4 antigen (CD24; CD24 molecule); ATPase type 13A2 (ATP13A2); epoxide hydrolase 1, microsomal (xenobiotic) (EPHX1); HtrA serine peptidase 1 (HTRA1); leptin receptor (LEPR); spectrin beta non-erythrocytic 1 (SPTBN1); muscleblind-like 2 (MBNL2); olfactory receptor family 2 subfamily J member 3 (OR2J3); Ras homolog enriched in brain (RHEB); glutamate receptor, ionotropic, N-methyl D-aspartate-associated protein 1 (GRINA); D-box binding protein, promyelocytic leukemia (PML), potassium inwardly-rectifying channel, subfamily J, member 2 (KCNJ2), topoisomerase (DNA) 1 (TOP1) and combinations thereof.

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